## Notes

# Silver nanoparticle modified Pt electrode as voltammetric and electrochemical impedance sensor for hydrogen peroxide in live biological cells

Kangkana Deka<sup>a</sup>, Jutika Kumar<sup>a</sup>, Ananya Bhowmick<sup>b</sup>, Sofia Banu<sup>b</sup> & Diganta Kumar Das<sup>a, \*</sup>

<sup>a</sup>Department of Chemistry, Gauhati University, Guwahati 781 014, Assam, India

Email: diganta\_chem@gauhati.ac.in

<sup>b</sup>Department of Bio-engineering and Technology, Institute of Science and Technology, Gauhati University, Guwahati 781 014, Assam, India

A non-enzymatic electrochemical sensor for hydrogen peroxide based on silver nanoparticle (AgNP) modified platinum electrode has been fabricated by multiple cyclic voltammetric scan of platinum electrode in AgNP solution in aqueous medium. The modified electrode (Pt/AgNP) can detect hydrogen peroxide in aqueous medium, bovine serum albumin and live L6 rat myoblast cells with high sensitivity and selectivity by cyclic voltammetry and electrochemical impedance spectroscopy. The limit of detection of Pt/AgNP towards  $H_2O_2$  is  $5.4 \times 10^{-7}$  M. The detection of  $H_2O_2$  by Pt/AgNP is free of interference from Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, dopamine and ascorbic acid.

Keywords: Electroanalytical chemistry, Sensors, Nanoparticles, Silver nanoparticles, Hydrogen peroxide, Cyclic voltammetry, Electrochemical impedance spectroscopy

Reactive oxygen species (ROS) are important intracellular signalling molecules, which participate in several physiological events such as protein synthesis, DNA damage, cell apoptosis, signal transition and immunity activity etc.<sup>1</sup> However, excess of ROS present in cells leads to various diseases, sometimes even causing death. Hydrogen peroxide  $(H_2O_2)$  is one of the common ROS found in different biological segments having several harmful effects. Therefore, the accurate detection and determination of  $H_2O_2$  in cells as well as in environment is very important in environmental, pharmaceutical, clinical and industrial research<sup>2,3</sup>. Among various analytical techniques such titrimetry, spectrophotometry, fluorescence, as chemiluminescence, and electrochemistry, the electrochemical sensing of H<sub>2</sub>O<sub>2</sub> is especially attractive due to low detection limit, high selectivity,

sensitivity, cost-effectiveness, fast response, high sensitivity, and ease of miniaturization<sup>4-10</sup>.

Enzyme based biosensors for H<sub>2</sub>O<sub>2</sub> have attracted considerable attention due to excellent substrate efficiency<sup>11-15</sup>. and high Poor specificity reproducibility, chemical and thermal instabilities and complicated immobilization procedures of these biosensors<sup>16,17</sup> have turned recent efforts of H<sub>2</sub>O<sub>2</sub> determination towards enzyme-free sensors<sup>18,19</sup>. Metal nanoparticles (NPs), due to their high electrical conductivity, high surface area and chemical stability, have become attractive electrode materials in electrochemical devices<sup>20</sup>. Metal NPs have become one of the popular and efficient electrode modifying agents for developing electrochemical sensors<sup>21</sup>. Electrode modified with metal NPs of Co<sup>22</sup>, Au<sup>23</sup>,  $Pt^{24}$ ,  $Pd^{25}$  and  $Ag^{26}$  are known for  $H_2O_2$  reduction.

In this work, we report a Pt electrode modified with the AgNPs synthesized from a reported method, which can detect  $H_2O_2$  by cyclic voltammetry, square wave voltammetry and electrochemical impedance spectroscopy. Dopamine (DA), ascorbic acid (AA), uric acid (UA) and various metal ions such as Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Cu<sup>2+</sup> and Zn<sup>2+</sup> do not interfere in the detection of  $H_2O_2$ . The modified electrode has been applied successfully for the detection of  $H_2O_2$  in live rat myoblast cells.

## Experimental

 $AgNO_3$  and hydrogen peroxide  $(H_2O_2)$  were purchased from Loba Chemie. Bovine serum albumin was purchased from Himedia. The electronic spectra were recorded on a UV-1800 Shimadzu spectrophotometer. SEM images were obtained on a Zeiss SEM analyzer. Electrochemical experiments were carried out on a CHI 660D electrochemical analyzer (USA) with a three-electrode cell system. All reagents were of analytical grade and used as received. The 0.1 M phosphate buffer solution (PBS) was prepared by mixing 19.0 mL of 0.1 M NaH<sub>2</sub>PO<sub>4</sub> and 81.0 mL of 0.1 M Na<sub>2</sub>HPO<sub>4</sub>.7H<sub>2</sub>O and was used as a supporting electrolyte in all electrochemical experiments. Fresh solution of H<sub>2</sub>O<sub>2</sub> was prepared experiment. before each All electrochemical experiments were carried out under N<sub>2</sub> environment. In the square wave voltammetry experiments, the square wave amplitude was kept as 25 mV, the frequency was 15 Hz and the potential height for base staircase wave front was 4 mV.

AgNPs were synthesized according to the reported procedure<sup>27</sup>. Briefly, 100 mL (1 mM) solution of AgNO<sub>3</sub> was taken in an Erlenmeyer flask and placed on a hot magnetic stirrer plate. *Azadirachta indica* plant leaves extract (1 mL) was added and the solution was stirred for 10 min. The colour of the solution changed from light yellow to brown supporting the formation of AgNP which was confirmed by UV/visible spectroscopy and scanning electron microscopic (SEM) studies.

AgNP modified Pt electrode was prepared as follows: Prior to the modification, Pt electrode was cleaned as per reported procedure<sup>28</sup> followed by sonication first in CH<sub>3</sub>OH and then in water. The electrode was dried under a stream of N<sub>2</sub> gas. The cleaned electrode was placed in the AgNPs solution containing 0.1 M NaNO<sub>3</sub> as the supporting electrolyte. Cyclic voltammetric runs were carried out for 100 scan segments at a scan rate 0.09 V s<sup>-1</sup>. The electrode was then gently washed with water and dried under a stream of N<sub>2</sub> gas and stored overnight in a refrigerator for use. The modified electrode is designated as AgNP/Pt henceforth in this report.

For the detection of  $H_2O_2$  in living Cells, rat L6 myoblasts cells were grown in DMEM medium supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin and maintained at 37 °C in a humidified atmosphere with 5% CO<sub>2</sub>. For the electrochemical study, the cells were seeded in a 6-well (35 mm) culture dish with a seeding density of  $3\times10^5$  cells per dish. After reaching 80% confluence, cells were washed with PBS (0.02 M, pH = 7.4) three times. Then 3 mL of PBS (0.02 M, pH = 7.4) was added to the plates for real sample measurements. The modified AgNP/Pt electrode, auxiliary and reference electrode were placed on the plate. Upon addition of AA to the system, the cells released H<sub>2</sub>O<sub>2</sub> which was measured electrochemically.

### **Results and discussion**

UV/visible spectrum of the aqueous solution of Cu NPs show a peak at 440 nm, which is the characteristic peak for AgNPs<sup>27</sup>. The morphology of the synthesized AgNPs was studied by scanning electron microscopy. Spherical nanoparticles of average size 100 nm were observed in SEM image (Supplementary data Fig. S1).

Figure 1 shows the cyclic voltammograms (100 scans) during the modification process of Pt electrode by Ag NPs. The increase in current indicates formation of AgNPs film on Pt electrode surface. The AgNP/Pt electrode was further characterised by recording double potential step chronocoulometry of AgNP/Pt in PBS containing 0.1 M NaNO<sub>3</sub> (Fig. 1, inset). The sharp decrease in charge versus time is indicative of the formation of electroactive film on the surface of Pt electrode.



Fig. 1 – Cyclic voltammograms of Pt electrode in Ag nanoparticle solution in water containing 0.1 M NaNO<sub>3</sub> as the supporting electrolyte for 100 scans at scan rate 0.09 V s<sup>-1</sup>. [Inset: Chronocoulogram of the modified electrode in water containing 0.1 M NaNO<sub>3</sub> as supporting electrolyte].



Fig. 2 – Cyclic voltammograms of AgNP/Pt at different scan rates in PBS containing 0.1 M NaNO<sub>3</sub>. [Inset: Plot of cathodic peak current versus scan rate].

The cyclic voltammograms of AgNP/Pt electrode in PBS at different scan rates are shown in Fig. 2. The reduction peak current was found to increase with scan rates. The plot of reduction current against scan rate was linear, confirming the formation of AgNPs film on the Pt electrode surface (Fig. 2, inset).

Figure 3 shows the cyclic voltammogram of AgNP/Pt in PBS at different added concentrations of  $H_2O_2$  (0.33–3.27 mM). In the absence of  $H_2O_2$ , an irreversible cyclic voltammogram was obtained with reduction peak at +0.048 V. Addition of  $H_2O_2$  shifted this reduction peak of AgNP/Pt to -0.035 V with



Fig. 3 – Cyclic voltammograms of AgNP/Pt in PBS containing 0.1 M NaNO<sub>3</sub> at different added concentration of  $H_2O_2$ . [Inset: Plot of the cathodic current versus  $H_2O_2$  concentration].



Fig. 4 – Cyclic voltammograms of AgNP/Pt in PBS containing 0.1 M NaNO<sub>3</sub> at different scan rates containing 3.27 mM  $H_2O_2$ . [Inset: Plot of cathodic current versus square root of scan rates].

gradual increase in the cathodic current. A good linear plot of the reduction peak current versus concentration of H<sub>2</sub>O<sub>2</sub> was observed with  $R^2 = 0.97$ . Cyclic voltammograms AgNPs/Pt was recorded at different scan rates in presence of 3.27 mM of H<sub>2</sub>O<sub>2</sub> (Fig. 4). The current was found to increase with the scan rate. The linearity of the cathodic current versus square root of scan rates plot ( $R^2 = 0.99$ ) indicates that the H<sub>2</sub>O<sub>2</sub> reduction process at AgNPs/Pt surface is a diffusion controlled process.

The mechanism of electrocatalytic reduction of  $H_2O_2$  by AgNP/Pt can be shown to follow  $C_iE_i$  mechanism as shown below. Here  $C_i$  stands for irreversible chemical reaction while  $E_i$  stands for irreversible electrochemical reaction.

$$AgNP + H_2O_2 + 2H^+ \longrightarrow Ag^+NP + H_2O \quad (Ci)$$
$$Ag^+NP + e^- \longrightarrow AgNP \quad (Ei)$$

The number of electrons involved in the overall reduction process of H<sub>2</sub>O<sub>2</sub> by AgNP/Pt can be calculated from the plot of current versus (scan rate) $^{1/2}$ . For an irreversible diffusion controlled process the relation between current (I) and scan rate (v) is given by  $I = 3.01 \times 10^5 \text{ n} [(1-\alpha)n_{\alpha}]^{1/2} \text{ACD}^{1/2} v^{1/2}$ . Here, diffusion co-efficient (D) was calculated as  $8.07 \times 10^{-6}$  cm<sup>2</sup> s<sup>-1</sup> with area of the electrode (A) as 0.91 cm<sup>2</sup> and concentration (C) of  $H_2O_2$  as  $0.34 \times 10^{-3}$  mol cm<sup>-3</sup>. [(1- $\alpha$ )n<sub> $\alpha$ </sub>] was considered as 0.7 as reported in literature<sup>1</sup>. The number of electrons involved (n) has been calculated as  $1.97 (\sim 2.0)$ . This value is in conformity with the value reported for electrocatalytic reduction of  $H_2O_2$ . The limit of detection (LOD) of H<sub>2</sub>O<sub>2</sub> was found as 0.541 µM and calculated by using the formulae. LOD = 3 (RSD/slope), where RSD = standard deviation for the average measurement of the blank sample and slope = calibration plot slope value<sup>29</sup>.

The H<sub>2</sub>O<sub>2</sub> sensing ability of AgNPs/Pt electrode was further studied by square wave voltammetry. Figure 5 shows the square wave voltammograms of AgNP/Pt electrode at different added concentration of H<sub>2</sub>O<sub>2</sub> in the electrolytic medium. With the addition of H<sub>2</sub>O<sub>2</sub> (0.33–3.27 mM) the peak current at potential +0.006 V increases from 3.909  $\mu$ A to 10.09  $\mu$ A. The relationship between peak current and concentration of H<sub>2</sub>O<sub>2</sub> is linear with  $R^2 = 0.98$ . The potential of AgNP/Pt electrode shifted from +0.058 V in absence of H<sub>2</sub>O<sub>2</sub> to +0.010 V when concentration of H<sub>2</sub>O<sub>2</sub> becomes 3.27 mM.



Fig. 5 – Square wave voltammograms of AgNP/Pt in PBS containing 0.1 M NaNO<sub>3</sub> at different added concentration of  $H_2O_2$ . [Inset: Plot of the cathodic current versus  $H_2O_2$  concentration].

The EIS measurements of interaction between AgNPs/Pt and H<sub>2</sub>O<sub>2</sub> were studied at  $E_{DC} = 1.5$  V. The EIS Nyquist plot of AgNP/Pt at different added concentration of H<sub>2</sub>O<sub>2</sub> in PBS shows the charge transfer resistance ( $R_{CT}$ ) to increase with increasing H<sub>2</sub>O<sub>2</sub> concentration (Fig. 6). The difference in charge transfer resistance ( $\Delta R_{CT}$ ) was found to increase linearly with increasing H<sub>2</sub>O<sub>2</sub> concentration (Fig. 6, Inset.

The selectivity of AgNPs/Pt electrode towards  $H_2O_2$  in the presence of biologically important metal ions and molecules has also been established. For this purpose, cyclic voltammograms of AgNP/Pt were recorded in PBS at different added concentration of  $H_2O_2$ , in presence of Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, DA and AA in the electrolytic medium. The nature of the cyclic voltammograms remained same and their peak positions were within ±0.010 V, as compared to those recorded without the above-mentioned interfering agents. This confirms the selectivity of AgNPs/Pt towards  $H_2O_2$  in the presence of Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, DA and AA.

Studies were carried out for the detection of  $H_2O_2$ in living cells and in bovine serum albumin (BSA): We detected current of  $H_2O_2$  released by the L6 rat myoblast cells lines stimulated by 1 µM AA. The cells were first washed with PBS solution (0.02 M; pH = 7.4) three times. AgNP/Pt in PBS (0.02 M, pH = 7.4) in presence of L6 rat myoblast cells did not show any specific current (Fig. 7, black line). Cyclic voltammogram recorded after addition of 1 µL AA to the cell solution showed the reduction peak



Fig. 6 – Nyquist plot of AgNP/Pt in PBS containing 0.1 M NaNO<sub>3</sub> at different added concentration of  $H_2O_2$ . [Inset: Plot of  $\Delta R_{CT}$  versus  $H_2O_2$  concentration].



Fig. 7 - Cyclic voltammograms of AgNP/Pt in rat L6 myoblasts cells (1), rat L6 myoblasts cells+AA (2), rat L6 myoblasts cells+AA+catalase (3).

corresponding to  $H_2O_2$  (Fig. 7, red line). In order to verify whether the current response after addition of AA was due to the  $H_2O_2$  produced from the living cells or not, 40 µL of catalase was added into PBS and cyclic voltammogram was recorded. The reduction peak was absent due to the metabolism of  $H_2O_2$  by catalase to water and oxygen (Fig. 7, blue line). These results suggest that the reduction current peak observed was only due to  $H_2O_2$  released from the living cells.

In order to investigate any damage to the cells due to the release of  $H_2O_2$  the optical morphology of cells before and after the addition of AA. The optical microscopy during the measurements showed that all the cells kept their regular shapes and there were no difference before and after the addition of AA (Supplementary data, Fig. S2).

The AgNPs/Pt electrode was also tested to detect  $H_2O_2$ in aqueous solution of bovine serum albumin (BSA).



Fig. 8 – Nyquist plot of AgNP/Pt in bovine serum albumin in PBS containing 0.1 M NaNO<sub>3</sub> at different added concentration of  $H_2O_2$ . [Inset: Plot of  $\Delta R_{CT}$  versus  $H_2O_2$  concentration].

Figure 8 shows the Nyquist plots of AgNPs/Pt when  $H_2O_2$  was added in the BSA medium up to 3.27 mM. The plot of the difference in charge transfer resistance ( $\Delta R_{CT}$ ) in the absence of  $H_2O_2$  in BSA medium is shown in Fig. 8 (Inset). A linear plot was obtained similar to that for aqueous medium.

In summary, AgNPs were prepared and characterized with the help of UV/visible spectroscopy and SEM analysis. The modified AgNPs/Pt electrode acts as an electrochemical sensor for  $H_2O_2$  by cyclic voltammetry, square wave voltammetry and electrical impedence spectroscopy. The AgNPs/Pt electrode can detect  $H_2O_2$  in presence of Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, DA and AA in aqueous medium. This electrode can also detect  $H_2O_2$  in living cells and in BSA-water medium by CV, SWV and EIS.

### Supplementary data

Supplementary data associated with this article are available in the electronic form at http://www.niscair.res.in/jinfo/ijca/IJCA\_57A(04)485-489\_SupplData.pdf.

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