

Indian Journal of Experimental Biology Vol. 60, April 2022, pp. 286-292



Synergistic effect of folic acid and galantamine against experimentally induced oxidative stress in IMR 32 cells

Shagun Dubey Upadhyay¹, Yusra Ahmad², Seema Kohli¹ & RK Sharma³

 ¹Department of Pharmacy, Government Kalaniketan Polytechnic College, Jabalpur, Madhya Pradesh, India
² Faculty of Pharmacy, Uttarakhand Technical University. Dehradun, Uttarakhand, India
³College of Veterinary Science & Animal Husbandry, Nanaji Deshmukh University of Veterinary Science, Jabalpur, Madhya Pradesh, India

Received 06 September 2021; Revised 24 January 2022

Galantamine is an active constituent obtained from Galanthus nivalis L., a traditional herb known for its pharmacological properties, particularly nootropic effect. Folic acid is a dietary supplement that enhances neuronal activity. Effect of galantamine and folic acid on human neuronal cells is well known. In the present study, we explored the protective effect of galantamine and folic acid, both independently as well as in combination, over antioxidant defence system and nootropic effects on human neuroblastoma cells IMR-32. The treatment galantamine, folic acid and their combination was given for 24 h and cytotoxicity study was carried out by trypan blue dye exclusion assay. Apoptosis and necrosis were observed using Propidium iodide (PI) and Hoechst double staining method. Biochemical assays viz. total protein, protein carbonyl, lipid peroxidation and glutathione were analyzed along with super oxide dismutase and catalase. Result of cytotoxicity showed dose dependent increase in percent viability and significant decrease was observed in apoptosis and necrosis. Moreover, exposure to Galantamine, Folic acid and their combination significantly decreased lipid peroxidation and protein carbonyl formation along with the enhancement in antioxidant defence mechanism. Findings of these dose reliant toxicity study of Galantamine, Folic acid and their combination suggest that these has higher potency when given together and shows synergistic effect. They also causes repair of human neuronal cells IMR-32 cells enhancing the cell viability and consumption of Galantamine and Folic acid together will help in prevention of CNS disorders and neurodegeneration.

Keywords: Galanthus nivalis, Nootropic, Neuroprotection, Snowdrop

Neurodegenerative diseases are one of the major health challenges especially in elderly population. It includes numerous diseases categorized by progressive loss of

*Correspondence:

E-Mail; dubeyshagun25@gmail.com

neurons. Various genetic and environmental factors have been found to modulate the risk for neurodegeneration¹. Neurodegeneration brings various challenges in the day to day life of an individual leading to difficulty in the survivals of the individual several nootropic agents have shown their effect on neurodegeneration. Nootropic agents, also known as smart drugs are highly acknowledged supplements that boost the cognitive performance. Nootropics increase the mental ability like learning, memory, attention, etc.².

Galantamine hydrobromide is a tertiary alkaloid which belongs to the *Amaryllidaceae* family². It is isolated from many species including *Leucojum* species, *Narcissus* species and *Galanthus* species. It is an important therapeutic option in various diseases³, including nerve pain and poliomyelitis⁴. It is a clinically approved drug for the treatment of Alzheimer disease which acts as a CNS-AChE inhibitor and allosteric potentiating ligand of the neuronal cholinergic nicotinic receptors^{6,7}. With the route of improvement, it has emerged as a massive beneficial preference in different neurological degeneration. Galantamine has been a clinically accepted drug for the remedy of Alzheimer's sickness. The medicine half way is going approximately as an AChE inhibitor that isan expected cholinergic nicotinic receptors⁸.

Folic acid is a dietary supplement that possesses fundamental roles in CNS and related functions in all age groups⁹. It is an inexpensive and multifunctional component that has shown its benefits in prevention of CNS disorders¹⁰. It also enhances the cellular neuronal differentiation by enabling biomechanical and biochemical pathway¹¹. Folic acid has also shown promising effects in functional neuronal recovery¹². It also possesses neuroprotective function as it inhibits the neuronal apoptosis via microRNAs¹³. In the current study, we tried to evaluate the neuroprotective effect of galantamine and folic acid on H₂O₂ and glutathione induced cytotoxicity in IMR-32 cells, which mimic cerebral cortex for better biological correlation.

Materials and Methods

Chemicals

All chemicals were procured from Hi Media, Mumbai, India and Sigma-Aldrich, USA.

Phone: + 91 8989203524 (Mob.)

Cell culture and Treatment

IMR-32 cell line was obtained from National Center for Cell Science (NCCS), Pune, India. The Cell line was maintained in Minimal Essential Media (MEM) supplemented with 2 mM L-glutamine, 0.1 mM non-essential amino acids and 10% Fetal bovine serum (HiMedia, Mumbai, India) in CO₂ incubator at 37° C with 5% CO₂.

Cytotoxicity testing by Trypan Blue dye exclusion assay

In our previous study, results of MTT assay showed that LC_{50} of galantamine and folic acid after 24 h of treatment to IMR-32 cells was 7 mM¹⁴. Based on this data, three different doses (2.5, 5 and 10 mM) of galantamine and folic acid, respectively were selected to evaluate the cytotoxic effects on human neuroblastoma cell line IMR-32 by trypan blue dye exclusion assay¹⁵.

Apoptosis and Necrosis observation by Propidium iodide (PI) and Hoechst 33342 Double staining

Cultures of IMR-32 cells were set up on coverslips for different experimental groups (Table 1) and after 24 h of galantamine and folic acid treatment (2.5, 5 and 10 mM) apoptosis and necrosis were observed using PI- 1 μ L (10 mg/mL) and Hoechst 33342- 1 μ L (10 mg/mL). Cells were examined under fluorescence microscope¹⁶ and the percentage of apoptotic and necrotic cells were calculated^{17,18}.

Percentage of Apoptotic Cells = $LA+DA/LN+LA+DN+DA \times 100$ and Percentage of Necrotic Cells = $DN/LN+LA+DN+DA \times 100$ where, LN= live cells with normal nuclei (PI/Hoechst 33342: blue chromatin with organized structure); LA= live cells with apoptotic nuclei (PI/ Hoechst 33342: bright blue chromatin that is highly condensed or fragmented); DN= dead cells with normal nuclei (PI/ Hoechst 33342: pink chromatin with organized structure); and DA= dead cells with apoptotic nuclei (PI/ Hoechst 33342: bright pink chromatin that is highly condensed or fragmented).

Oxidative stress indices

IMR-32 cells were seeded in 12-Well plates (10^{5} cells/Well) and cultured for 24 h. After 24 h of galantamine, folic acid and galantamine + folic acid treatment respectively, these cells were used to make cell lysate. The cells were trypsinized and treated with lysis buffer (pH 7.5) containing 1% Triton X-100, 130 mM NaCl, 10 mMTris-HCl and 10 mM NaH₂PO₄¹⁹. The mixture was incubated for 30 min at 4°C. The supernatant was used for biochemical assays like total protein (TP)²⁰, protein carbonyl (PC)²¹, lipid

peroxidation $(LPO)^{22}$ and total glutathione $(GSH)^{23}$ along with the activity of superoxide dismutase $(SOD)^{24}$ and catalase $(CAT)^{25}$.

Results

Cell viability study

The effect of galantamine on cell viability was significant trypan blue dye exclusion assay. The exposure of IMR-32 cells at lower doses galantamine was somewhat similar to the control group i.e. 89.9% cell viability. At increasing dose (5 mM), it was exactly similar to the control group i.e., 90.6%, live cells. When compared to the higher dose of 10 mM galantamine, the cell mortality was decreased up to 5%. (Fig. 1) whereas, in the case of folic acid, the results were remarkable. The exposure to IMR-32 cells at lower doses folic acid near to the control group i.e. 90.3% cell viability. When the dose was increased i.e. 5 mM, it was unerringly comparable to the control group i.e., 94.6%, live cells. When compared to the higher dose of 10 mM folic acid, the cell mortality was decreased up to 5.7%. (Fig. 1)

While studying the synergistic effect of galantamine and folic acid, the results were impressive. The exposure of IMR-32 cells at lower doses of galantamine and folic acid showed 90.5% cell viability. When the dose was increased to 5 mM it increased to 91% of live cells. At higher dose of 10 mM, a significant decrease in cell mortality was observed up to 7%. Thus, significant synergism was obseerved in the IMR32 cell line by galantamine and folic acid.

Apoptosis and Necrosis Observation

Galantamine and Folic acid treatment with lower dose i.e. 2.5 mM showed non-variant decrease in apoptosis, whereas at higher doses i.e. 5 and 10 mM,



Fig. 1 —Effect on Live and Dead cells after trypan blue dye exclusion assay in control (Gr. 1); and Galantamine (Gr. 2, 5, 8); Folic acid (Gr. 3, 6, 9); and Galantamine+Folic acid (Gr. 4, 7, 10) treated groups.

respectively showed potential decrease (P < 0.001) in apoptosis in association to the control group. Whereas the effect of galantamine and folic acid with all the three doses (2.5, 5 and 10 mM) showed effective decrease (P < 0.001) in necrosis when compared to control group. In the study of synergism of galantamine + folic acid at lower dose i.e. 2.5 nM, there was nonremarkable decrease in apoptosis while at higher doses i.e. 5 and 10 mM, respectively the decrease was significant (P < 0.001) in apoptosis when compared to the control group. In the case of effect of galantamine +folic acid on necrosis the, the decrease was note worthy with all the three doses (P < 0.001) in comparison to the control group (Table 1).

Oxidative stress analysis

Galantamine and Folic acid treatment showed an increase in total protein significantly at mid and higher doses (P < 0.05 and P < 0.01, respectively) whereas non-significant alteration was observed at low dose treatment as compared to control (Fig. 2A). Protein carbonyl can be formed inside the cell due to the breakage of protein backbone by generation of ROS or direct oxidation of amino acids and it showed no significant change at low dose of galantamine and folic acid treatment. At higher dose of galantamine and folic acid treatment the results where somewhere close to normal (P < 0.001) as there was no breakage of protein (Fig. 2B). In order to determine the level of malondialdehyde, the level of lipid peroxidation (LPO) was measured and it showed no change at low doses (2.5 mM) of galantamine and folic acid treatment. On the other hand the mid and high doses of galantamine and folic acid treatment (Group 8 and 9) showed promising decrease (P < 0.001) in LPO as compared to control (Fig. 2C). Glutathione is a major defence molecule against ROS. The level of GSH was observed

Table 1 — Percentage of apoptosis and necrosis by Propidium		
iodide and Hoechst double staining method		
Groups	Apoptosis	Necrosis
Control (Gr. 1)	6.3 ± 0.66	13.1±1.2
Galantamine low dose (2.5 mM) (Gr. 2)	6.6 ± 0.37	13.36 ± 1.5
Folic Acid low dose (2.5 mM) (Gr. 3)	$6.0{\pm}1.3$	11.2 ± 1.6
Galantamine+ Folic acid low	5.8 ± 1.4	12.1±2.2
dose(2.5+2.5 mM) (Gr. 4)		
Galantamine mid dose (5 mM) (Gr. 5)	$1.9{\pm}0.17$	12.02 ± 2.1
Folic acid mid dose (5 mM) (Gr. 6)	5.7 ± 0.2	11.9±1.1
Galantamine+Folic acid mid dose	4.9 ± 1.1	10.7 ± 1.3
(5+5 mM) (Gr. 7)		
Galantamine high dose (10 mM) (Gr. 8)	4.1 ± 1.0	8.7±1.3
Folic acid high dose (10 mM) (Gr. 9)	$4.0{\pm}1.9$	8.8 ± 1.2
Galantamine+Folic acid high dose	$3.9{\pm}0.9$	$7.9{\pm}1.0$
(10+10 mM) (Gr. 10)		

to be increasing significantly (P < 0.05, P < 0.01 and P < 0.001), respectively for 2.5, 5 and 10 mM doses of galantamine and folic acid treatment in dose dependent manner as compared to control. These parameters showed an excellent linear dose response relationship in cultured IMR-32 cells after 24 h of galantamine and folic acid exposure (Fig. 2D). Superoxide dismutase is an enzyme that catalysis the dismutation of O2- into oxygen and H_2O_2 whereas, catalase converts H_2O_2 to non-toxic water molecule. The SOD activity was non prominent in any groups (P < 0.001) in comparison to control (Fig. 2E). The CAT activity was noticed to moderately increase (P < 0.01) for the group of cells treated with 10 mM galantamine and folic acid treatment (Group 10) (Fig. 2F). While studying the synergism of galantamine + folic acid value of total protein was high in mid and higher doses (P < 0.05 and P < 0.01, respectively (Fig. 2A). Protein carbonyl was low as there was no breakage of protein (Fig. 2B). The lipid peroxidation (LPO) level was apparently negligible at any dose (Fig. 2C). The GSH level showed significant increase (Fig. 2D). The SOD activity in galantamine + folic acid was absolutely non variant in any groups (P < 0.001) in comparison to control (Fig 6). The activity of CAT was increased notably (P < 0.01) for the group of cells treated with 10 mM galantamine and folic acid treatment (Group 10) (Fig. 2F).

Discussion

Neurodegenerative disorders are varied group of diseases affecting the nervous system, including the brain, spinal cord and peripheral nerves that have different etiologies²⁶. Due to the prevalence, morbidity and mortality of the neurodegenerative diseases, they represent significant medical, social and financial burden to the society²⁷. Galantamine is a phytoconstituent that acts as a neuroprotective agent. Galantamine is a natural alkaloid having antioxidant properties. Galantamine is an inhibitor of acetylcholinesterase and has allosteric actions on nicotinic receptors. It is a scavenger of reactive oxygen species and exerts neuroprotection mainly by inhibition of the oxidative damage. It also alters the level of glutamate level in the brain that is involved in destruction of the neurons²⁸. Folic acid is the most important dietary supplement that is well studied for protective effect on neuronal function²⁹. It has also shown beneficial role in CNS and related activities³⁰. It is multifunctional component that has shown its benefits in prevention of CNS disorders³¹. It enhances



Fig. 2 — Effect on the Levels of (A) Total protein; (B) Protein carbonyl; (C) Lipid peroxidation; (D) GSH; (E) SOD; and (F) CAT in control (Gr. 1); and Galantamine (Gr. 2, 5, 8); Folic acid (Gr. 3, 6, 9); and Galantamine+Folic acid (Gr. 4, 7, 10) treated groups at low, mild and high dose with @2, 5, 10 mM, respectively [Values are expressed as mean \pm SEM (P < 0.01) and (P < 0.001)]

the cellular neuronal differentiation by enabling biomechanical and biochemical pathway³².

Folic acid has also shown its notable effects in functional neuronal recovery³³. It also possesses neuroprotective function as it inhibits the neuronal apoptosis via micro RNAs³⁴.Folic acid is an essential source of the single carbon group used in DNA methylation and plays a pivotal role in the development, function, regeneration, and repair of the central nervous system (CNS)³⁵. In this study, the level of glutamate was found to be elevated in cultured human neuroblastoma cell line IMR-32 studied using

MTT assay. The cell viability was negligible and the cell mortality was decreased up to 5% in case of galantamine treated group whereas it was 5.6% in folic acid treated group but even more promising result was seen in case of galantamine + folic acid treated group i.e. up to 7% which exclaimed the synergistic neuroprotective and neuroregenerative effect. The ratio of live and dead cells clearly showed that treatment of galantamine, folic acid and galantamine + folic acid treated group increased cell viability decreasing the rate of mortality. Moreover, propidium iodide and Hoechst 33342 double staining results revealed that

galantamine, folic acid and galantamine + folic acid treated group reduces apoptosis as well as necrosis in dose dependent manner. Excitotoxicity exerted by glutamate is also responsible for increase in the cytosolic Ca^{2+} level, which is due to either influx from the extracellular space or release from the intracellular stores³⁶. In this condition the survival of a cell depends largely on functioning of the mitochondria³⁷.

As mitochondria not only satisfy the cellular energy demands but also involved in ROS generation, which in turn are suspected to cause cell death if they get out of control³⁸. Increased ROS level causes damage to cell in several ways and oxidation of macromolecules like lipid, proteins and DNA. Increase in lipid peroxidation level due to ROS production results in loss of function and integrity of neuronal cell membranes, which in turn results in increase in non-specific permeability to ions, leading to disruption of membrane structure and cell functions³⁹. The malondialdehyde, a marker of lipid peroxidation was decreased significantly in galantamine + folic acid treated group, as well as in galantamine and folic acid treated groups. Oxidation of proteins and amino acids induced by ROS generation also resulted in increase of protein carbonyl level and these could be determined by carbonyl groups (aldehydes and ketones) that are produced on protein side chains when they are oxidized⁴⁰. In this study, a decrease in protein carbonyl level with corresponding increase in total protein confirms the notable rise in neuronal protein content. Antioxidant defense system consist mainly glutathione (GSH), superoxide dismutase (SOD) and catalase (CAT) as they are protagonist in our body⁴¹. The combination of Galantamine and folic acid protects the alteration of the antioxidant defense system and reduces oxidative stress. GSH protects the cells against free radical peroxides and other toxic compounds 42 .

In current study, dose dependent increment of GSH level was found after galantamine and folic acid treatment. Superoxide dismutase defends the cell against free radical injury by converting O_2 - radical to hydrogen peroxide (H₂O₂) and prevents the formation of OH– radicals through O_2 - driven Fenton reactions⁴³. The H₂O₂ formed by SOD is removed by catalase. Hence, if the activity of CAT is not adequate to degrade H₂O₂ into H₂O and O₂, than more H₂O₂ is converted into toxic hydroxyl radicals and finally responsible for cellular damage^{44,45}. Here, galantamine with folic acid was found to increase GSH levels as well as adhere the SOD and CAT activities, ultimately leading to the inhibition of neuronal cell damage.

Conclusion

The above results have demonstrated significance of both galantamine and folic acid treatments in neuroprotection. Galantamine and folic acid combination has shown more promising results in neuroprotection and as well as oxidative stress as indicated by cell viability and decreased apoptotic and necrotic behaviour of the cells. Apparent changes in the levels of total protein, protein carbonyl, lipid peroxidation, GSH, SOD and CAT in the treated groups have further shown synergistic effect of increased neuroprotection and decreased damage to the cells caused by oxidative stress. The neuro-protective effect of galantamine and folic acid on H₂O₂ and glutathione induced cytotoxicity in IMR-32 cells have also been shown.

Conflict of interest

Authors declare no competing interests.

References

- 1 Akhtar A, Andleeb A, Waris TS, Bazzar M, Moradi AR, Awan NR & Yar, Neurodegenerative diseases and effective drug delivery: A review of challenges and novel therapeutics. *J Control Release*, 330 (2021) 1152.
- 2 Johnson J, Mercado-Ayon E, Mercado-Ayon Y, Dong YN, Halawani S, Ngaba L & Lynch DR, Mitochondrial dysfunction the development and progression of neurodegenerative diseases. *Arch Biochem Biophys*, 702 (2021) 108698.
- 3 Upadhyay SD, Ahmad Y, Kohli S & Sharma RK, Evaluation of acetylcholinesterase and butyrylcholinesterase inhibitory activity of Huperzine-A; *In silico* and *in vitro* studies. *Indian J Biochem Biophys*, 56 (2019) 224.
- 4 Ganeshpurkar A & Saluja A, The pharmacological potential of hesperidin. *Indian J Biochem Biophys*, 56 (2019)287.
- 5 Koola MM, Galantamine-Memantine combination in the treatment of Alzheimer's disease and beyond. *Psychiatry Res*, 293 (2020) 113409. doi: 10.1016/j.psychres.2020.113409.
- 6 Metz CN & Pavlov VA, Treating disorders across the lifespan by modulating cholinergic signaling with galantamine. *J Neurochem*, 158 (2021) 1359.
- 7 Moreta MP, Burgos-Alonso N, Torrecilla M, Marco-Contelles J & Bruzos-Cidón C, Efficacy of Acetylcholinesterase nhibitors on Cognitive Function in Alzheimer's Disease. *Rev Res Biomeds*, 9 (2021) 1689.
- 8 Kowal NM, Ahring PK, Liao VW, Indurti DC, Harvey BS, O'Connor SM, Chebib M, Olafsdottir ES & Balle, Galantamine is not a positive allosteric modulator of human α4β2 or α7 nicotinic acetylcholine receptors. *Br J Pharmacol*, 175 (2018) 2911.
- 9 Bohra BS & Kale PP, Evaluation of nootropic effects of galantamine and sildenafil as a combination in mice. *Indian J Exp Biol*, 56 (2019) 60.
- 10 Roffman JL, Neuroprotective effects of prenatal folic acid supplementation: why timing matters. *JAMA Psychiatry*, 75 (2018) 747.
- 11 Yilmaz M., Aktug H., Oltulu F & Erbas O, Neuroprotective effects of folic acid on experimental diabetic peripheral neuropathy. *Toxicol Ind Health*, 32 (2016) 832.

- 12 Davis CK, Nampoothiri SS & Rajanikant GK, Folic acid exerts post-ischemic neuroprotection *in vitro* through HIF-1α stabilization. *Mol Neubio*, 55 (2018) 8328.
- 13 Budni J, Molz S, Dal-Cim T, Martín-de-Saavedra MD, Egea J, Lopéz MG, Tasca CI & Rodrigues ALS, Folic acid protects against glutamate-induced excitotoxicity in hippocampal slices through a mechanism that implicates inhibition of GSK-3β and iNOS. *Mol Neubio*, 55 (2018)1580.
- 14 Polavarapu S, Dwarakanath BS, & Das UN, Arachidonic acid activates extrinsic apoptotic pathway to enhance tumoricidal action of bleomycin against IMR-32 cells. *Prostaglandins Leukot. Essent. Fatty Acids*, 132 (2018) 16.
- 15 Shah N, Nariya A, Pathan A, Patel A, Chettiar SS & Jhala D, Neuroprotection effects of Celastruspaniculatus seed oil against monosodium glutamate in human IMR-32 cells. *Annu. Res Rev Biol*, 24 (2018) 1-9.
- 16 Tasneem S & Yasmeen R, Biochemical alterations in total proteins and related enzymes in tissues of Cyprinus. *Indian J Exp Biol*, 59 (2021), 125.
- 17 Samanta P & Ghosh AR, Evaluation of health status of Oreochromis niloticus (L.) based on brain optic tectum morphopathology. Indian J Exp Biol, 59 (2021) 877.
- 18 Raut GK, Manchineela S, Chakrabarti M, Bhukya CK, Naini R, Venkateshwari A, Reddy VD, Mendonza JJ, Suresh Y, Nallari P & Bhadra MP, Imine stilbene analog ameliorate isoproterenol-induced cardiac hypertrophy and hydrogen peroxide-induced apoptosis. *Free Radic Biol Med*, 153 (2020) 80.
- 19 Gurram B, Li M, Li M, Gebremedhin KH, Sun W, Fan J, Wang J & Peng X, NIR-excited superoxide radical procreators to eradicate tumors by targeting the lyso-membrane. *J Mater Chem B*,7 (2019) 4440.
- 20 Thakur AK & Movileanu L, Real-time measurement of protein–protein interactions at single-molecule resolution using a biological nanopore. *Nat Biotechnol*, 37 (2019) 96.
- 21 Mano JI, Biswas M & Sugimoto K, Reactive carbonyl species: a missing link in ROS signalling. *Plant*, 8 (2019) 391.
- 22 Bhatt D, Vyas K, Singh S, John PJ & Soni I, Tartrazine induced neurobiochemical alterations in rat brain sub-regions. *Food Chem Toxicol*, 113 (2018) 322.
- 23 Boeing T, Mariano LN, Dos Santos AC, Tolentino B, Vargas AC, de Souza P, Nesello LA & da Silva LM, Gastroprotective effect of the alkaloid boldine: Involvement of non-protein sulfhydryl groups, prostanoids and reduction on oxidative stress. *Chem Biol Intera*, 25 (2020) 327.
- 24 Kumar RR, Dubey K, Goswami S, Hasija S, Pandey R, Singh PK, Singh B, Sareen S, Rai GK, Singh GP & Singh AK, Heterologous expression and characterization of novel manganese superoxide dismutase (Mn-SOD)–A potential biochemical marker for heat stress-tolerance in wheat (Triticum aestivum). *Int J Bio Macromol*, 161 (2020) 1029.
- 25 Hadwan MH & kadhum Ali S, New spectrophotometric assay for assessments of catalase activity in biological samples. *Anal Biochem*, 542 (2018) 29.
- 26 Ransohoff RM, How neuroinflammation contributes to neurodegeneration. *Science*, 353 (2016) 777.
- 27 Poprac Patrik KlaudiaJomova, Miriama Simunkova, Vojtech Kollar, Christopher J. Rhodes & Marian Valko, Targeting free

radicals in oxidative stress-related human diseases. *Trends Pharmacol Sci*, 38 (2017) 592.

- 28 Yang J & Kim G, Local delivery of epigenetically stimulating folic acid for peripheral nerve regeneration. 42nd Society for Biomaterials Annual Meeting and Exposition 2019: The Pinnacle of Biomaterials Innovation and Excellence. Society for Biomaterials.
- 29 Stewart KJ, Iskandar BJ, Meier BM, Rizk EB, Hariharan N, Koueik J, Andrei AC & Hogan KJ, Nitrous oxide impairs axon regeneration after nervous system injury in male rats. *Anesth*, 131(2019) 1063.
- 30 Fernández-Villa D, Jiménez Gómez-Lavín M, Abradelo C, San Román J & Rojo L, Tissue engineering therapies based on folic acid and other vitamin B derivatives. Functional mechanisms and current applications in regenerative medicine. *Int. J Mol Sci*, 19 (2018) 4068.
- 31 Kang WB, Chen YJ, Lu DY & Yan JZ, Folic acid contributes to peripheral nerve injury repair by promoting Schwann cell proliferation, migration, and secretion of nerve growth factor. *Neural Regen Res*, (2019) 132.
- 32 Balashova, O.A., Visina, O. & Borodinsky, L.N, Folate action in nervous system development and disease. *Dev Neurobiol*, 78 (2018) 391.
- 33 Sağır D, Kuruoğlu E, Onger ME & Yarar E, The Effects of folic acid and topiramate on peripheral nerve regeneration. *Biotech Histochem*, (2019) 1.
- 34 Iskandar BJ, Nelson A, Resnick D, Pate Skene JH, Gao P, Johnson C, Cook TD & Hariharan N, Folic acid supplementation enhances repair of the adult central nervous system. *Annals of Neurology*, 56 (2004) 221.
- 35 Mezeiova E, Spilovska K, Nepovimova E, Gorecki L, Soukup O, Dolezal R, Malinak D, Janockova J, Jun D, Kuca K & Korabecny J, Profiling donepezil template into multipotent hybrids with antioxidant properties. *J Enzyme Inhib Med Ch*, 33 (2018) 583.
- 36 Reggiani AM, Simoni E, Caporaso R, Meunier J, Keller E, Maurice T, Minarini A, Rosini M & Cavalli A, In vivo characterization of ARN14140, a memantine/galantaminebased multi-target compound for Alzheimer's disease. *Sci Rep*, 6 (2016) 33172.
- 37 Dar NJ, Bhat JA, Satti NK, Sharma PR, Hamid A & Ahmad, Withanone an active constituent from Withania somnifera, affords protection against NMDA-induced excitotoxicity in neuron-like cells. *Mol Neurobiol*, 54 (2017) 5061.
- 38 Shah N, Singh R, Sarangi U, Saxena N, Chaudhary A, Kaur G, Kaul SC & Wadhwa R, Combinations of Ashwagandha leaf extracts protect brain-derived cells against oxidative stress and induce differentiation. *PLoS One*, 10 (2015) 0120554.
- 39 Hazman M, Hause B, Eiche E, Nick P & Riemann M, Increased tolerance to salt stress in OPDA-deficient rice ALLENE OXIDE CYCLASE mutants is linked to an increased ROS-scavenging activity. J Exp Bot, 66 (2015) 3339.
- 40 Davies MJ, Protein oxidation and peroxidation. *Biochem J*, 473 (2016) 805.
- 41 Ighodaro OM & Akinloye OA, First line defence antioxidantssuperoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): Their fundamental role in the entire antioxidant defence grid. *Alexandria J Med*, 54(2018) 287.
- 42 Gaschler MM & Stockwell BR, Lipid peroxidation in cell death. Biochem Biophys Res Commu, 482 (2017) 419.

- 43 Smirnoff N & Arnaud D, Hydrogen peroxide metabolism and functions in plants. *New Phytologist*, 221 (2019) 1197.
- 44 Mishra P & Sharma P, Superoxide Dismutases (SODs) and their role in regulating abiotic stress induced oxidative stress in plants. In: *Reactive Oxygen, Nitrogen and Sulfur Species in Plants: Production, Metabolism, Signaling and*

Defense Mechanisms, (Eds. Hasanuzzaman M, Fotopoulos V, Nahar K & Fujita M; John Wiley & Sons Ltd., NJ, USA), 2019 53,88.

45 Nandi A, Yan LJ, Jana CK & Das N, Role of catalase in oxidative stress-and age-associated degenerative diseases. Oxid Med Cell Longev, (2019).