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Cassava starch film loaded with extract of *Piper betel* leaf for anti-inflammatory activity

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The aim of the present research work was to formulate a transdermal film of *Piper betel* L. leaf extract along with diclofenac sodium and evaluate its anti-inflammatory activity. The ethanolic extract of *P. betel* leaf was obtained by solvent extraction method using a Soxhlet apparatus. The *P. betel* leaf extract was analysed for inhibition of the albumin denaturation and it showed a significant inhibition of the albumin denaturation. The topical film was then prepared by solvent casting method using diclofenac sodium and *P. betel* leaf extract. The film was evaluated for drug content, thickness, weight uniformity, folding endurance, moisture content, moisture uptake, and barrier properties of the film. The formulated film was thin, flexible, and possessed satisfactory physicochemical properties. The anti-inflammatory potential of the film was evaluated by using carrageenan-induced rat paw oedema method on albino Wistar rats. The anti-inflammatory activity of film was found comparable with standard diclofenac sodium film and exhibited inhibition of oedema within 4 h. This signifies that the combination of *P. betel* leaf extract and diclofenac sodium has remarkable potential to serve as synergistic topical anti-inflammatory film preparation than simple topical film formulation of diclofenac sodium.

Keywords: Anti-inflammatory activity, Cassava starch, Diclofenac sodium, Piper betel, Transdermal film.

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Introduction

Drug delivery by the topical route is an emerging and convenient method. The topical route has variety of advantages compared to oral route such as avoidance of first pass metabolism, ease of administration, convenient, helps to achieve control and sustained release of drug in plasma and better patient compliance¹. The topical application of the agent generally refers to delivery of the drug through the skin either for systemic or for local action. The transdermal delivery of drug in the form of film provides the advantage of avoiding dose dumping and in case, if any toxicity develops, it allows the patient to terminate the drug therapy by simply removing the film^{1,2}. In response to various stimuli such as pathogens, damaged cells and irritants, body initiates its defence mechanism called as inflammation. There is an increased blood flow to the area of injury or infection due to the release of chemicals and results in pain, redness, and swelling in the affected area. In

response to injury or infection, the immune system triggers this physical reaction. The inflammatory response is provoked by the production of mediators and chemotactic factors³⁻⁵. Non-steroidal antiinflammatory drugs (NSAIDs) are generally preferred for the treatment of inflammation. The oral dosage form of the NSAID undergoes hepatic first-pass metabolism and only half of the administered dose reaches the systemic circulation⁶⁻⁸. On prolonged usages it causes ulcer, increases gastric irritation, nausea, and vomiting. NSAIDs have severe side effects such as delay in muscle regeneration, ulceration, haemorrhage, tissue damage, etc. and on prolonged use cause gastrointestinal bleeding, heart attack, stroke, high blood pressure, and kidney damage⁸. Due to these, disadvantages, there is a need to search for an alternative to overcome the problems associated with the use of NSAIDS⁹⁻¹¹. Nowadays, interest is increasing day by day towards natural remedies with a basic approach towards nature. The dependency on the use of herbal or traditional medicine has increased from all over the world with approximately 80% of the world population inclined towards the use of herbal medicine to cater to their primary health care needs.

P. betel L. is cultivated in India, Sri Lanka, Malaysia, Indonesia, and East Africa. Its extracts are reported to cure urinary tract infections, cervicitis, vaginitis, gastrointestinal disorders, skin infections, etc^{12,13}. Currently, there are abundant scientific findings reporting the beneficial effects of betel leaves including antioxidant, anti-carcinogenic, antiinflammatory, antibacterial, antifungal, and antidiabetic activities¹²⁻¹⁷.

Cassava starch is a polysaccharide and is considered as one of the most promising natural biopolymers. The films developed from Cassava starch are easily biodegradable, tasteless, odourless, colourless, and non-toxic¹⁸⁻²¹.

Considering the anti-inflammatory properties of P. betel leaf extracts, an attempt was made to develop a transdermal film of P. betel leaf extract and diclofenac sodium. As per the literature review, this is the first work reported to combine the extract of P. Betel and the synthetic drug diclofenac sodium with an intention that this combination would reduce the dose of the diclofenac sodium, which ultimately could help to reduce the side effects associated with the use of NSAIDS (diclofenac sodium) for a longer duration of time along with the benefits of topical route. The research work of combining P. betel leaf extract and diclofenac sodium is based on the hypothesis that, if the individual constituent exerts similar effect, then their combination will sometimes exhibit enhanced effect. The synergistic effect achieved could allow the use of lower doses of the active pharmaceutical ingredient and help to reduce adverse effects related to their prolonged use²².

Materials and Methods

Cassava starch was obtained from Shree Ram Bio Starch Polymers Private Limited, Vadodara, India. All other chemicals used in project work were of analytical grade.

Plant collection and identification

P. Betel leaves were collected in the month of July 2018. The identification was done in the Department of Botany, SGPKM, Girad, Wardha, where a voucher specimen (ATNPIPERB-04) was deposited. The leaves, stems, and roots were separated and cut into small pieces. They were dried in an oven at 40 °C to constant weight. The drying time and percentage of water loss were determined in the process.

Preparation of Betel leaf extract

The leaves of *P. betel* were washed with distilled water, dried in shade and crushed into a fine powder. The dried powder was extracted using ethanol (1:15) using a Soxhlet apparatus for 24 h at 60 °C. The extract was dried in an oven for 3 days at 40 °C until the entire solvent was evaporated until the residue was left. The dried extract was weighed, recorded and stored in a tightly closed container^{15,21}.

Phytochemical screening of plant extract

To obtain ethanolic extract of *P. betel*, few drops of different reagents were added in an individual test tube such as 5% ferric chloride solution, lead acetate solution, bromine water, potassium dichromate, dilute iodine solution, dilute nitric to determine the phenolic group present in the plant extract¹⁴.

Inhibition of albumin denaturation

To investigate the anti-inflammatory activity of the *P. betel* leaf extract, a method suggested by Sakat *et al.* was followed with minor modifications²³. The ethanolic extracts of *P. betel* leaf (sample) and diclofenac sodium (control) at different concentration ranging from 10-80 µg/mL were mixed with 1% aqueous solution of bovine albumin. To this mixture, a small amount of 1N hydrochloric acid solution was added to adjust the pH. The samples were then kept in the incubator for 20 min at 37 °C. Thereafter, the samples were heated to 57 °C for 20 min. After cooling, the turbidity produced was determined spectrophotometrically at 660 nm. The percent inhibition of protein denaturation was then calculated using the following equation:

Percentage inhibition of protein denaturation (%)

$$=\frac{Abs_{control} - Abs_{sample}}{Abs_{control}} X 100\%$$

Fourier transform infrared spectroscopy

The FTIR spectra of *P. betel* leaf extract, diclofenac sodium, and the formulated medicated film of (*P. betel* leaf extract + diclofenac sodium) were recorded using FTIR spectrophotometer (Shimadzu) using the single reflection horizontal attenuated total reflectance (ATR) to determine drug excipient compatibility. The FTIR spectrum was recorded from 4000 to 390 cm^{-1(Ref. 24,25)} (Fig. 1a-d).

Film preparation

Initially, Cassava starch 4 g was added into distilled water and was boiled at 90 °C to carry out the gelatinization of starch. While boiling, the solution



Fig. 1 — FTIR Spectrum, a) *Piper betel* leaf, b) Diclofenac sodium, c) Cassava starch, d) Test film (*Piper betel* leaf extract and Diclofenac sodium).

was stirred continuously until homogeneous. Thereafter, glycerine 2 mL was added into that solution. This mixture was cooled at 40-45 °C. Then, the ethanolic extract of *P. betel* leaf (40 mg) and diclofenac sodium (100 mg) was added into this mixture. The mixture was then transferred into the Petri plate and kept in the oven for drying for 24 h. After drying, the film formed in Petri plate was peeled off carefully²⁶.

Physicochemical characterization of the film

The formulated film was evaluated for the following physicochemical characterization methods

Thickness

Digital micrometre screw gauge was used to measure the thickness of the film. The thickness was measured at three different places of film and the mean value was calculated²¹.

Weight variation

The weight variation was studied individually for each film²⁶.

Folding endurance

The numbers of times the film could be folded at the same place without breaking denotes folding endurance. The film was repeatedly folded at the same place until it broke.

Drug content

To measure the drug content, 2×2 cm² size of the film was kept in a phosphate buffer pH 5.5 and shaken continuously for 24 h. The solution was ultrasonicated for 15 min. After filtration, the drug (diclofenac sodium) content was estimated spectrophotometrically at λ_{max} of 278 nm.

Moisture uptake

The film was weighed and kept in a desiccator at room temperature for 24 h. The film was then exposed to 84% relative humidity (a saturated solution of aluminium chloride) and was weighed again. The percentage of moisture uptake was calculated as the difference between final and initial weight with respect to initial weight as shown in following equation:n

Moisture uptake(%) = $\frac{W \text{ final} - W \text{ initial}}{W \text{ initial}} \ge 100$

Moisture content

The films were weighed individually and dried in an oven at 105 °C for 24 h. After removal of the film from the oven, the films were weighed again and their weights were recorded. The percentage of moisture content was calculated as the difference between initial and final weight with respect to final weight as shown in following equation:

Moisture content (%) =
$$\frac{W \text{ initial} - W \text{ final}}{W \text{ final}} \ge 100$$

Barrier properties of the film

Sterile nutrient broth solution about 10 mL was poured into two clean test tubes. Cassava starch film (2 cm^2) loaded with *P. betel* leaf and diclofenac sodium extract was applied to one test tube, while the other test tube was kept uncovered. Both the test tubes were kept aside for 24 h in the incubator and the growth of microorganisms was observed after 24 h¹⁸.

In vivo studies

Healthy male Albino Wistar rats weighing 200-250 g were used in the study. The rats were maintained in the animal house of JSPM's Rajarshi Shahu College of Pharmacy and Research, with controlled temperature, 25±1 °C for 10-12 h light-dark cycle at 23 °C and provided with water and food. The experimental protocol was approved by IAEC (Institutional Animal Ethics Committee), JSPM's Rajarshi Shahu College of Pharmacy and Research, Tathawade, Pune protocol number RSCPR/IAEC/2019/05. Laboratory animal handling and the research protocol was performed in accordance with CPCSEA guideline²⁷⁻²⁹.

Carrageenan-induced rat paw oedema

The modified method of Winter *et al.* 1962; Otterness and Oore, 1988 was used in the present study to develop carrageenan–induced rat paw oedema³⁰⁻³². Twenty four male Wistar rats of 180-200 g were used and divided into four groups, each group consisting of six rats (n=6). Group I served as normal control group and received no treatment. Groups II served as the positive control received carrageenan injection (0.1 mL, 1% w/v in normal saline). Group III received transdermal film (containing 40 mg of P. betel leaf extract and 100 mg of diclofenac sodium) and served as treatment group. Group IV received diclofenac sodium patch (containing 200 mg of diclofenac sodium) and served as a standard group. Oedema was induced by injecting freshly prepared carrageenan (0.1 mL, 1% w/v in normal saline) into the sub-planter tissue of each right hind paw in all groups animal except normal control group. The paw circumference was measured before and at 1, 2, 3, and 4 h after carrageenan injection using screw gauze method. Both standard and test patch were applied on sub plantar tissue of right hind paw and the result of both were observed and compared with the positive control group. The percentage value of oedema inhibition was calculated by the following formula²³:

Percentage inhibition (%) =
$$1 - \frac{y-x}{b-a} X100$$

where, x=Initial paw thickness of test group animals, y=Paw thickness of test group animal after treatment, a=Initial paw thickness of control group animal, b=Paw thickness of control group animal after treatment.

Haematological parameters

After 4 h to carrageenan induction and after application of formulation film and standard film, blood samples were collected by retro-orbital route in heparin tube and submitted to a pathological laboratory in Pune. Haematological parameter namely white blood cells (W.B.C.) count was analyzed²⁹.

Statistical analysis

The Graph pad prism version 5 (Graph Pad Software Inc., La Jolla, CA) was used for statistical analyses which were presented as mean \pm SEM. The data were statistically analysed by two-way analysis of variance (ANOVA) followed by Bonferroni post hoc test, where *P* <0.01 was considered as statistically significant.



Fig. 2 — a) Injection of 1% carrageenan solution, b) Observation after injecting carrageenan solution, c) Application of test film, d) Removal of test film after 6 h.

Results and Discussion

Phytochemical analysis

In the present study, the preliminary phytochemical analysis of the ethanolic extract of P. betel leaf was carried out to detect the presence of the active constituent phenol. On addition of 5% ferric chloride solution into the extract, the colour of the solution changed from green to deep blue, lead acetate solution changed the colour of the extract from green and formed white precipitate. Bromine water caused the decolouration of the test solution; potassium dichromate changed the colour of nitric acid into the extract, the colour changed the colour of the extract from green to red; on addition of nitric acid into the extract, the colour changed from green to reddish yellow, thus indicating the presence of phenol in the extract.

Inhibition of albumin denaturation

The ethanolic extract of *P. betel* at a dose of 40 μ g/mL exhibited an anti-inflammatory activity as presented in Table 1. Protein denaturation is an indicator of inflammatory disease. The agents that can prevent protein denaturation can act as an anti-inflammatory agent. The anti-inflammatory effect of *P. betel* leaf extract was highest at a concentration of 40 μ g/mL.

Thickness and weight uniformity, drug content

All the films were found to be uniform in thickness, homogeneous with smooth surface without any sign of cracking. The average thickness of all formulated films were observed to be in the range of 0.1 mm, the weight of the film was in the range of 2.145-2.366 mg and the drug content was found in between 91.24-98.64%.

Folding endurance, tensile strength

The films exhibited good flexibility with sufficient mechanical properties. The result indicated that the film would not break and maintain their integrity when applied on the skin. Without cracks, the folding endurance value of the film was 130 when folded at

Table 1 — Percent Inhibition of albumin denaturation				
Concentration (µg/mL)	Inhibition of albumin denaturation (%)			
10	14.2 ± 0.3			
20	30±0.5			
30	30.43±0.2			
40	50±0.6			
50	38±0.1			
60	35±0.6			
70	40 ± 0.8			
80	38.46±0.1			

the same place, thus indicating good folding endurance.

Moisture uptake

All the film showed an increase in the moisture uptake with the time, with increase in the concentration of *P. betel* leaf extract, the percentage of moisture uptake also increased²¹. This could be because of the presence of the phenolic group in the extract.

Moisture content

The moisture content of the prepared formulation was low, which helps in maintaining its stability throughout long-term storage at dry condition, thereby preventing the film from drying and becoming brittle.

Barrier property of the film

No turbidity was observed in the test tube sealed with the film which indicated that the film acted as a barrier with antimicrobial properties preventing the entry of micro-organism into the nutrient broth, whereas the test tube which was uncovered exhibited turbidity due to the growth of micro-organism.

Anti-inflammatory activity

Carrageenan-induced rat paw oedema method is the most widely used and most accepted model for inflammation. Carrageenan induces inflammation by releasing different inflammatory mediators such as histamine, serotonin, and bradykinin. In the present study, carrageenan was used to induce inflammation in all the groups except normal control Group I. Carrageenan was injected in the sub-plantar region of paw circumference of rats in Group II, Group III, and Group IV²⁸⁻³⁰. Each group was observed 4 h after the injection of carrageenan. Also, the effect of carrageenan on the rat paw oedema was observed after the application of the standard marketed film of diclofenac sodium and the test film (P. Betel extract and diclofenac sodium). The result for the antiinflammatory activity was predicted in terms of per cent inhibition of paw circumference. In the normal control group, the paw circumference was 0.374±0.04 mm. In Group II which received carrageenan showed significant injection а increase in paw circumference as compared to control Group I (no carrageenan was injected). Group III also exhibited an increase in paw circumference before application of test film. After treatment with the test film, the paw circumference was reduced (Fig. 2 a-d). In the group III, after the injection of carrageenan, the

readings recorded for the decrease in the paw circumference at the time interval of 1, 2, 3, 4 h were 0.698 ± 0.08 , 0.547 ± 0.02 , 0.511 ± 0.02 , 0.0483 ± 0.01 respectively and % inhibition of paw circumference at 1, 2, 3, 4 h were 38.12, 51.50, 54.69, 57.18% respectively for the groups considered for the study. In comparison to Group II (no treatment), the rats in the Group III, which received the test film showed less significant (P < 0.05) decrease in paw circumference at 2^{nd} and 3^{rd} h interval, while it showed significant decrease in paw circumference at 4th h and was found to be statistically significant (P < 0.001). Group IV showed an increase in paw circumference before application of the standard film, which then found to decrease after the application of the standard film containing (diclofenac sodium). The paw circumference reading after the application of the film at 1, 2, 3, and 4 h were 1.118±0.04, 0.874±0.006, 0.553±0.018, 0.518±0.025, 0.479±0.033 respectively and % inhibition of paw circumference at 1, 2, 3, 4 h were 21.82, 50, 53.66, 57.15% respectively (Fig. 3). After the injection of Corrageenan, the increase in the rat paw circumference as shown in Fig. 3 was significantly reduced in Group III and Group IV. The per cent inhibition of the paw circumference of rats in Group III and Group IV showed a significant decrease in



Fig. 3 — Carrageenan induced paw oedema.

paw circumference at 3rd and 4th h interval as shown in Table 2. This indicates that the combination of diclofenac sodium (100 mg) with the P. betel leaf extract showed comparable anti-inflammatory activity when compared with the standard diclofenac sodium film containing 200 mg of diclofenac sodium. The results indicated that combination of P. betel leaf extract with diclofenac sodium (100 mg) had positive impact on the anti-inflammatory activity of diclofenac sodium and exhibited same anti-inflammatory activity as observed with diclofenac sodium 200 mg. Thus, the combination of P. betel leaf extract with diclofenac sodium resulted in reducing the dose of diclofenac sodium to almost 50%. The hypothesis by the authors suggesting the combination of P. betel leaf extract and diclofenac sodium can be supported by research work by various researchers. Nitric oxide (NO) is an important signalling molecule which is produced at the inflammation site by the action of NO synthase present in leucocytes. NO is a proinflammatory mediator known to induce inflammation³¹. P. betel leaf contains polyphenols which are reported to be the potent inhibitors of nitric oxide synthase and nitric oxide production 32,33,34 . P. betel leaf extract also possess free radical and peroxynitrite (ONOO) (-) scavenging activity³⁵. The findings by Alam B et al. suggested that P. betel may be used as a supplementary herbal remedy for the treatment of pain and inflammatory disease³⁶. The authors further proposed that the analgesic and antiinflammatory effect exerted by P. betel extract may prove to be valuable when combined with analgesic and anti-inflammatory drugs³⁶. Earlier studies have reported that the P. betel leaf extract exhibited synthesis^{37,38}. cyclooxygenase inhibition of Cyclooxygenase is known to initiate the formation of prostaglandins and increased the level of prostaglandins in the tissue initiates pain, oedema, and

Table 2 — Percent inhibition of paw circumference in different groups of rats						
Groups	Before application		Paw size in mm at time in h. and % inhibition			
	of film Paw circumference	1h	2h	3h	4h	
Group I	$0.373 {\pm} 0.04$	$0.373 {\pm} 0.04$	$3.73 {\pm} 0.004$	$3.73 {\pm} 0.04$	$3.73 \pm .004$	
% inhibition of paw circumference	0%	0%	0%	0%	0%	
Group II	1.123±0.05***	$0.850{\pm}0.009^{***}$	$0.668 {\pm} 0.009 {***}$	$0.659 \pm 0.009 ***$	$0.632 \pm 0.008 ***$	
% inhibition of paw circumference	0%	24%	40.51%	42.11%	43.72%	
Group III	1.128 ± 0.04	$0.698 \pm 0.08 **$	$0.547 \pm 0.02*$	$0.511 \pm 0.02*$	$0.483 \pm 0.01 **$	
% inhibition of paw circumference	0%	38.12%	51.50%	54.69%	57.18%	
Group IV	1.118 ± 0.04	$0.874 {\pm} 0.006$	$0.553 {\pm} 0.018$	$0.518 \pm 0.025*$	$0.479 \pm 0.033 **$	
% inhibition of paw circumference	0%	21.82%	50%	53.66%	57.15%	



fever. Inhibitory effect of the *P. betel* leaf extract on cyclooxygenase is comparable to that produced by NSAIDs. These NSAIDs also acts by similar mechanism by inhibiting cyclooxygenase enzyme. Hence, the combination of *P. betel* leaf extract and diclofenac sodium would prove to be beneficial as an anti-inflammatory agent. The result of this research work supports the hypothesis of combining *P. betel* leaf extract with NSAIDs to improve the anti-inflammatory activity and reduce the dose of diclofenac sodium by exerting synergistic effect.

This approach of combining *P. betel* leaf extract with diclofenac sodium would reduce the load on the synthetic drug thus providing an alternative to reduce the side effects associated with the use of synthetic agents.

Haematological parameter

The result for the haematological parameter is given in Fig. 4. The W.B.C. count in the Control group I was found to be 9200 cu/mm³. In Group II, the WBC count was increased to 19000 cu/mm³ after injection of carrageenan. In Group III, after the injection of carrageenan, rise in W.B.C. count was observed to 166000 cu/mm³ which was found to decrease after the application of test film (P. betel leaf extract and diclofenac sodium film) to 8900 cu/mm³. In Group IV standard film i.e., diclofenac sodium patch treated group showed an increase in W.B.C. count after injection of carrageenan solution to 13200 cu/mm³ which was reduced to 7000 cu/mm³ after application of the standard film. The total WBC count for an adult ranges from 5,000 to 10,000/mm in a normal condition³⁹. WBC > 10,000/cu/mm³ represents inflammation (possibly due to allergies) and tissue damage⁴⁰. Carrageenan, a chemical obtained from red algae, stimulates the release of inflammatory and pro-inflammatory mediators. Cytokines are a series of pro-inflammatory mediators activated by macrophages and are capable to stimulate WBC by enhancing the attachment and migration of WBC's on the endothelial cells, thereby increasing their levels in the blood as observed in Group II indicating the development of inflammation⁴¹. In Group III rats treated with the test film a decline in the WBC count was observed due to anti-inflammatory effect of P. betel leaf extract. It can suppress the secretion of proinflammatory cytokines responsible to stimulate WBC^{42,43}. The decline in WBC count in Group III was almost comparable to that observed in Group IV. The reason for this might be the combination of *P. betel* leaf extract and diclofenac sodium which must have resulted in decline in the WBC count which was evident even in the dose of 100 mg of diclofenac sodium and almost comparable to that observed in group IV, wherein the dose of diclofenac sodium was 200 mg. The results of haematological parameter also support the hypothesis of combining P. betel leaf extract with NSAIDs to improve the antiinflammatory activity and reduce the dose of diclofenac sodium.

Conclusion

The study indicates combination of P. betel leaf extract with diclofenac sodium would prove to be beneficial in improving the anti-inflammatory activity of diclofenac sodium. The study suggested that the combination of diclofenac sodium (100 mg) with P. betel leaf extract exhibited the same anti-inflammatory activity as that of diclofenac sodium (200 mg). This in turn would prove to be beneficial to reduce the untoward effects associated with the use of higher dose of NSAIDs, thereby improving patient compliance.

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Conflict of interest

Authors declared 'no conflict of interest'.

References

- 1 Jeong W Y, Kwon M, Choi H E and Kim K S, Recent advances in transdermal drug delivery systems: A review, *Biomater Res*, 2021, **25**(1), 1-15.
- 2 Priyanka K, Kumar G and Uttam S B, Formulation optimization and characterization of transdermal film of curcumin by response surface methodology, *Chin Herb Med*, 2021, **13**(2), 274-285.

- 3 Juhn S K, Jung M K, Hoffman M D, Drew B R, Preciado D A, *et al.*, The role of inflammatory mediators in the pathogenesis of otitis media and sequelae, *Clin Exp Otorhinolaryngol*, 2008, 1(3), 117-138.
- 4 Abdulkhaleq L A, Assi M A, Abdullah R, Zamri S M, Taufiq-Yap Y H, *et al.*, The crucial roles of inflammatory mediators in inflammation: A review, *Vet World*, 2018, 11(5), 627-635.
- 5 Sugimoto M A, Sousa L P, Pinho V, Perretti M, and Teixeira M M, Resolution of inflammation: What controls its onset?, *Front Immunol*, 2016, 7, 1-18.
- 6 Alqahtani M S, Kazi M, Alsenaidy M A and Ahmad M Z, Advances in oral drug delivery, *Front Pharmacol*, 2021, 12, 1-21.
- 7 Sinha M, Gautam L, Shukla P K, Kaur P, Sharma S, *et al.*, Current perspectives in NSAID-induced gastropathy, *Mediat Inflamm*, 2013, **2013**, 1-11.
- 8 Wongrakpanich S, Wongrakpanich A, Melhado K and Rangaswami J, A comprehensive review of non-steroidal anti-inflammatory drug use in the elderly, *Aging Dis*, 2018, 9(1), 143-150.
- 9 Bindu S, Mazumder S and Bandyopadhyay U, Non-steroidal anti-inflammatory drugs (NSAIDs) and organ damage: A current perspective, *Biochem Pharmacol*, 2020, **180**, 1-21.
- 10 Nunes C D R, Arantes M B, Pereira S M F, Cruz L L, Passos M S, et al., Plants as sources of anti-inflammatory agents, *Molecules*, 2020, 25(16), 1-22.
- 11 Maroon J C, Bost J W and Maroon A, Natural anti-inflammatory agents for pain relief, *Surg Neurol Int*, 2010, **1**, 1-10.
- 12 Syahidah A, Saad C R, Hassan M D, Rukayadi Y, Norazian M H, et al., Phytochemical analysis, identification and quantification of antibacterial active compounds in betel leaves, *Piper betle* methanolic extract, *Pak J Biol Sci*, 2017, 20(2), 70-81.
- 13 Rintu D, Shinjini M, Kaustab M, Pramathadhip P, Umesh P S, et al., Anti-oxidant and anti-inflammatory activities of different varieties of Piper leaf extracts (*Piper betle L.*), J Nutr Food Sci, 2015, 5, 1-16.
- 14 Satish A B, Deepa R V, Rohan V G, Nikhil C T, Yatin Y R, et al., Phytochemistry, pharmacological profile and therapeutic uses of *Piper betle* Linn-An overview, *Int J Pharmacogn Phytochem Res*, 2013, 1(2), 10-19.
- 15 Hoque M M, Rattila S, Shshir M A, Bari M L, Inatsu Y, et al., Antibacterial activity of ethanol extract of betel leaf (*Piper betle* L) against some foodborne pathogen, Bangladesh J Microbial, 2011, 28(2), 58-63.
- 16 Toprani R and Patel D, Betel leaf: Revisiting the benefits of an ancient Indian herb, South Asian J Cancer, 2013, 2(3), 140-141.
- 17 Nayaka N M D M W, Sasadara M M V, Sanjaya D A, Yuda P E S K, Dewi N L K A A, et al., Piper betle (L): Recent review of antibacterial and antifungal properties, safety profiles, and commercial applications, *Molecules*, 2021, 26(8), 1-21.
- 18 Souza A C, Goto G E O, Mainardi J A, Coelho A C V and Tadini C C, Cassava starch composite films incorporated with cinnamon essential oil; Antimicrobial activity, microstructure, mechanical and barrier properties, *J Food Sci Technol*, 2013, 54, 346-352.
- 19 Medina J C, Gutiérrez T J, Goyanes S, Bernal C and Famá L, Biodegradability and plasticizing effect of yerba mate extract

on cassava starch edible films, *Carbohydr Polym*, 2016, **20**, 150-159.

- 20 Chinma C E, Ariahu C C and Alakli J S, Effect of temperature and relative humidity on the water vapour permeability and mechanical properties of cassava starch and soy protein concentrate based edible films, *J Food Sci Technol*, 2013, **52**(4), 2380-2386.
- 21 Leila N and Abdorreza M N, Antibacterial, mechanical and barrier properties of sago starch film incorporated with betel leaf extract, *Int J Biol Macromol*, 2014, **66**, 254–259.
- 22 Lehár J, Krueger A S, Avery W, Heilbut A M, Johansen L M, *et al.*, Synergistic drug combinations tend to improve therapeutically relevant selectivity, *Nat Biotechnol*, 2009, 27(7), 659-668.
- 23 Sakat S S, Juvekar A R and Gambhire M N, *In-vitro* antioxidant and anti-inflammatory activity of methanol extract of *Oxalis corniculata* Linn, *Int J Pharm Pharm Sci*, 2010, 2, 146–55
- 24 Khairi M S F, Abdullah S, Ramez M and Sadik S, First derivative ATR-FTIR spectroscopic method as a green tool for the quantitative determination of Diclofenac sodium tablets, *F1000Res*, 2020, **9**, 1-14.
- 25 Singh T P, Chauhan G, Agrawal R K and Mendiratta S K, In vitro study on antimicrobial, antioxidant, FT-IR and GC– MS/MS analysis of Piper betle L. leaves extracts, J Food Meas Charact, 2019, 13(1), 466-475.
- 26 Ahmad N, Tayyeb D, Ali I, Alruwaili K, Ahmad N, *et al.*, Development and characterization of hemicellulose-based films for antibacterial wound-dressing application, *Polymers*, 2020, **12**(3), 548.
- 27 Anyasor G A and Ijituyi O H, Formulated hexane fraction of costus after leaves balm suppressed xylene induced topical inflammation in rat model, *Am J Physiol Biochem Pharmacol*, 2018, 7(2), 54-60.
- 28 Gupta R and Gupta G D, Formulation development and evaluation of anti-inflammatory potential of *Cordia oblique* Topical gel on Animal model, *Pharmacogn J*, 2017, 9(6s), s93-s98.
- 29 Kshirsagar A D, Panchal P V, Harle U N, Nanda R K and Shaikh H M, Anti-inflammatory and antiarthitic activity of anthaquinone derivatives in rodent, *Int J Inflam*, 2014, 2014, 1-12.
- 30 Winter C A, Risley E A and Nuss G W, Carrageenin induced oedema in bind paw of the rat as assay for anti-inflammatory drugs, *Exp Biol Med*, 1962, **111**, 544–547.
- 31 Otterness I G and Moore P F, Carrageenan foot oedema test, *Methods Enzymol*, 1988, **162**, 320- 327.
- 32 Mansouri M T, Hemmati A A, Naghizadeh B, Mard S A, Rezaie A, et al., A study of the mechanisms underlying the anti-inflammatory effect of ellagic acid in carrageenaninduced paw edema in rats, *Indian J Pharmacol*, 2015, 47(3), 292-298.
- 33 Sharma J N, Al-Omran A and Parvathy S S, Role of nitric oxide in inflammatory diseases. *Inflammopharmacol*, 2007, 15(6), 252–259.
- 34 Srivastava R C, Husain M M, Hasan S K and Athar M, Green tea polyphenols and tannic acid act as potent inhibitors of phorbol ester induced nitric oxide generation in rat hepatocytes independent of their antioxidant properties, *Cancer Lett*, 2000, 153(1–2), 1-5

- 35 Viana G S B, Bandeira M A M and Matos F J A, Analgesic and anti-inflammatory effects of chalcones isolated form Myracrodruon urundeuva Allemao, *Phytomed*, 2003, 10(2-3), 189–195
- 36 Alam B, Akter F, Parvin N, Pia R S, Akter S, et al., Antioxidant, analgesic and anti-inflammatory activities of the methanolic extract of *Piper betle* leaves, *Avicenna J Phytomed*, 2013, 3(2), 112-125.
- 37 Gupta M, Mazumder U K, Gomathi P and Selvan V T, Antiinflammatory evaluation of leaves of *Plumeria* acuminate, BMC Complement Altern Med, 2006, 6(1), 1-6.
- 38 Sawadogo W R, Boly R, Lompo M, Some N, Lamien C E, et al., Anti-inflammatory, analgesic and antipyretic activities of Dicliptera verticillata, Int J Pharmacol, 2006, 2(4),435–438.
- 39 George E L and Panos A, Does a high WBC count signal infection?, *Nursing*, 2005, **35**(1), 20-21.

- 40 Blumenreich M S, The white blood cell and differential count, In *Clinical Methods: The History, Physical, and Laboratory Examinations*, edited by H K Walker, W D Hall, J W Hurst, 3rd edn, (Boston: Butterworths), 1990.
- 41 Davydova V N, Sorokina I V, Volod'ko A V, Sokolova E V, Borisova M S, *et al.*, The comparative immunotropic activity of carrageenan, chitosan and their complexes, *Mar Drugs*, 2020, **18**, 458.
- 42 Suprapto M, Cytotoxicity of betel leaf (*Piper betel* L.) against primary culture of chicken embryo fibroblast and its effects on the production of proinflammatory cytokines by human peripheral blood mononuclear cells, *Dent J*, 2012, 45(2), 97–101.
- 43 Arango D G and Descoteaux A, Macrophage cytokines: Involvement in immunity and infectious diseases, *Front Immunol*, 2014, 5, 491.