

Indian Journal of Natural Products and Resources Vol. 13(4), December 2022, pp. 419-433 DOI: 10.56042/ijnpr.v13i4.47930



A review of the nutritional potential of edible snails: A sustainable underutilized food resource

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Received 24 March 2021; revised received 04 November 2022; accepted 24 November 2022

Despite the use of snails as food in many different communities in the world, the consumption of snail meat is still far from being appreciated. The authors compiled and reviewed the nutrient composition of edible snail meat based on the available scientific literature. The protein, fat, and ash content of snail meat were found to be 14.0, 1.4 and 2.1% on the basis of fresh matter respectively. Comparable protein content with conventional foods of animal origin along with the presence of all essential amino acids and less carbon footprint makes the snail meat a sustainable food source. Moreover, less fat content but higher proportion of polyunsaturated fatty acids, and considerable presence of minerals with nutritional importance especially calcium, zinc, and iron could exhibit human nutritional benefits. However, the establishment of heliciculture facilities is a necessity in order to support this sustainable food resource and also enhance the economic condition of certain local areas.

Keywords: Amino acids, Angiostrongyliasis, Food safety, Food security, Minerals, Protein IPC code; Int. cl. (2021.01)-A23J, A61K 36/00

Introduction

According to the recent report of the global hunger index¹, 828 million people are suffering from undernourishment (defined as a lack of sufficient calories in the food) and inadequate nutrition (defined as a lack of nutrients in the diet). Sub-Saharan Africa and South Asia are regions with the highest hunger and undernutrition levels in the world¹. At least 2 billion people are suffering from micronutrient deficiency which is often termed "the hidden hunger"², but at the same time about 2 billion people suffer from overweight or obesity. In the context of a expanding global population still and the unanticipated climate change, we as humans are forced to explore alternative sustainable food sources to feed the people in future.

We need to increase the production of nutrient-rich food without harming the environment or increasing greenhouse gas emissions, or demanding to provide additional water, but with an aim to achieve low feed conversion ratios and to intensify farming systems

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without laying off farm workers. To understand the linkages between sustainability, productivity or yield of food not only the unit area of land has to be considered but inputs like irrigation needs, fertilizers, insecticides, herbicides, machinery, fuel costs, emission of greenhouse gases (GHG) etc. must not be overlooked either. It was first suggested by Meyer-Rochow to use unconventional (alternative) foods, like for instance edible insects to ease global food shortages and to involve the FAO as well as WHO³. The idea slowly gained acceptance and has led to the current interest in entomophagy⁴. Algae as a food source has also received some attention in the last few decades⁵, but snails as a candidate to augment the spectrum of available food items has been given a comparatively poor attention in the agriculture sector. This is somewhat surprising because a variety of snail species are even now commonly consumed by members of numerous communities around the world. In fact, the use of snails as food has a long history and in the Mediterranean region has been known from prehistoric times⁶ to this day⁷.

According to the estimates of FAOSTAT⁸, there is an increasing trend to produce food based on snails (marine

species not included) year by year (Fig. 1). On average, 15% of the snails come from snail-breeding units, while the remaining 85% are harvested from Nature⁹. Snail farming, known as 'Heliciculture', is not at all a new entrant in the agriculture sector, but initiatives to explore this food resource have been limited and the resource clearly remains as under-utilized.

In the era of globalization, we tend to lose traditional foods with the result of nutrient deficiencies and negative health consequences¹⁰. Most often the interruption of a regular supply plays a critical role in the declination of traditional foods. Halwart, reported that there was no tradition of snail farming and marketing in south-east Asia although rural people of different parts of this region preferred to eat native snails¹¹. In Japan, snail farming became popular in the 90s, and many governmental and nongovernmental livelihood projects were established to support a backyard cottage industry¹¹. Snails have also been used as feed for broiler chickens, ducks, and pigs¹¹⁻¹³. However, because of an assumed high pest potential, farming systems did not continue to use snails much as animal feed and, to cite an example, Vietnam banned snail farming in mid-1992. In many parts of India, local people do collect edible terrestrial as well aquatic snails from the forest and water bodies and consume them¹⁴ (personal observations of SG). Recently some small scale snail farms have been developed (personal communication of SG).

In a survey on bushmeat in Nigeria, Martin placed snails in a separate category from bushmeat as they were very common (especially the giant African snail Archachatina marginata) particularly in the forested areas where people did not consider them as bushmeat¹⁵. For the last few decades Africa, and in particular western Africa, is experiencing a rapid growth of the number of snail farms with the adequate help of governmental as well as non-governmental agencies¹⁶⁻¹⁸. Snail farming has also received attention in developed country like Korea. Ghosh et al., described the farming system of Pomacea canaliculata, as edible snail, as it is practiced as minilivestock in Korea¹⁹. From 1995 to 2010, Italian snail production and consumption increased almost four times and the annual sale in 2013 was worth about 265 million Euros²⁰. Allen estimated that the consumption of edible snail Theba pisana accounts for approximately 4000 tons per year^{7,21}. Snail farming is characterized by not involving labourintensive work, or requiring high capital investment, sophisticated technical and professional or knowledge, and this makes it suitable for developing and/or underdeveloped areas. However, in order to appreciate the nutritional advantages of the rearing and consumption of snails, it is essential to examine and highlight the nutrient composition of the snails. The primary aim of this review has therefore been to evaluate the nutrient content of the edible species.



Fig. 1 — Global production of snails (marine species not included) with the year (Source: FAOSTAT, 2021).

Edible snail species in the world

Based on the available scientific literature, the authors compiled an inventory of edible snails around the globe and presented it in Table 1. Although there are undoubtedly many more, we focused on those species that are widely popular and have, to some extent, been scientifically studied with regard to chemical composition, biology, and breeding. Such

	able 1 — Inventory of edible snail and bivalve species aroun		
Scientific name	Geographical location	Purpose	Reference
Africa			
Helix aspersa	Morocco	Food	50
Limicolaria aurora	Uyo main market, Akwa Ibom state, Nigeria	Food	51
Archachatina marginata	Ikere-Ekiti market, Ondo state, Nigeria	Food	52
Archatina			
Limicolaria			
Achatina achatina	Ghana	Food	53
<i>Limicolaria</i> sp.	Oba market (Ado-Ekiti), Ekiti state, Nigeria	Food	54
Archatina archatina			
Archachatin amarginata			
Lymnaea stagnalis	Kuto market in Abeokuta, Ogun state, Nigeria	Food	55
Archachatina marginata ovum	King's market, Ado Ekiti, Ekiti state, Nigeria	Food	56
Archachatina marginatasaturalis	King 5 market, 146 Ekiti, Ekit Suite, 16gena	1000	50
Achatina achatina			
Limicolaria sp.			
Archachatina marginata ovum	Wild collection from Calabar, Odukpani and Ugep;	Food	57
Archachatina marginatasaturalis		1000	51
	Nigeria		
<i>Limicolaria</i> sp.			
Lanistes varicus			
Nucella lapillus		F 1	-0
Archachatina marginata	Markets in Ibadan metropolis, Oyo state, Nigeria	Food	58
Achatina achatina			
Achatina fulica			
Achatina fulica	Uyo, Akwa Ibom state, Nigeria	Food	59
<i>Limicolaria</i> sp.			
Helix pomatia			
Helix trunculus	Sfax city fish market, Tunisia	Food	60
Archachatina marginata	Minna, Niger state, Nigeria	Food	61
Archachatina marginata	Farm at Rivers State University of Education,	Food	62
	Ndele campus, Nigeria		
Achatina achatina	Farm in Kumasi, Ghana	Food	63
Limicolari flammea	Agboville, Côte d'Ivoire	Food	64
Archachatina marginata	Itoku market, Abeokuta, Ogun state, Nigeria	Food	65
	Oje market, Oyo state, Nigeria		
	Oyingbo market, Lagos state, Nigeria		
	Oja Oba market, Osogbo, Osun state, Nigeria		
	Oja Oba market, Akure, Ondo state, Nigeria		
Archachatina marginata	Oyo state, Nigeria	Food	66
Archachatha marginala Achatina fulica	Littoral (Wouri), Center (Lekie) and West (Santchou),	Food	67
		roou	0 /
Archachatina marginata	Cameroon		
	Asia	E 1	(0
Pila scutata	Malaysia	Food	68
Bellamyiaingallsiana	T 11 1 1 1 TZ	F 1	10
Ampullarius insularus	Jepllabuk-do, Korea	Food	69
Achatina fulica			
Pomacea canaliculata	Thailand	Food	11
	Vietnam		
	Taiwan		
	Philippines		
Achatina fulica	Junchun Food Co., Kimhae, Korea	Food	70
Pila globosa	29 tribal groups in Bangladesh	Food	71,72
Apple snail	Japan	Food	73
Helix pomatia	Local processing company in Cukurova, Turkey	Food	74
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	1 — Inventory of edible snail and bivalve species around the	· · · · ·	D C
Scientific name	Geographical location	Purpose	Reference
Semisulcospira gottschei	Korea	Food	75
Achatina fulica	China	Food	76
Pomacea canaliculata		F 1	
Bellamya bengalensis	North East India	Food	77
Bellamya bengalensis f. typica			
Bellamya bengalensis f. balteata			
Bellamya dissimilis			
Pila globosa			
Pila scutata			
Pila theobaldi			
Pila virens			
Brotia costula	Northann word of Dillow (Khanania, Salama, Madhaman	T-41	22
Bellamya sp.	Northern part of Bihar (Khagaria, Saharsa, Medhepura,	Ethnomedicinal	23
Bellamya bengalensis	Purnea), India	uses	
Pila scutata			
Lamellidens sp.			
Parreysia sp.			
Lamellidens sp.	Varia	East	70
Semisulcospira coreana	Korea	Food	78
Semisulcospira gottschei			
Semisulcospira forticosta			
Semisulcospira libertina			
Semisulcospira tegulata		F 1	70
Helix aspersa	Asian Istanbul, Turkey	Food	79
Viviparid snail	Vietnam	Food	80,27
Chamberlainiahainesiana	Thailand, Vietnam	Food	01
Brotiacostula	Wild (pond, paddy field, wetland) and market,	Food	81
Bellamya bengalensis	Lembucherra, Tripura, India		
Bellamya dissimilis			
Pila globosa		E4 1'' 1	24
Helix aspersa	Tribal people of Chhindwara (Gond, Bharia, Mawasi,	Ethnomedicinal	24
	Ahir), Madhya Pradesh, India Tailad halt of Can have a sing Madhee	uses	25
Pila sp.	Tribal belt of Gondwanavindhyan region, Madhya	Ethnomedicinal	25
Pomacea canaliculata	Pradesh, India	uses Food	82
	Farm in Andong, Korea Consumed by Muda tribe, Jharkhand, India	Food	82 83
<i>Pila globosa</i> Unidentified snails species	Local markets (collected from wild areas like water	Food	Personal experience
Undentified shalls species	bodies) in northern part of West Bengal, India	rood	of SG
Unidentified analis analis	Consumed by Baiga tribe, Madhya Pradesh, India	Food	14
Unidentified snails species	Australia	rood	14
Cookia auloata		Food	84
Cookia sulcata	Te Oka Bay, Canterbury, New Zealand (consumed by	rood	04
Turbo militaris	Maori people) Woolgoolga, Coffs harbour, Australia	Food	85
Lunella undulata	woorgoorga, corrs naroour, Austrania	roou	00
Lunella torquata	Europa		
Holix adenarsa	Europe	Food and	22,86
Helix adspersa Helix pomatia	Europe		22,00
Helix pomatia Helix lucorum		Therapeutic	
		uses	
Helix aperta Helix cincta			
Otala sp.			
Eobania vermiculata	Form and wild Latoria	Food	07
Helix pomatia	Farm and wild, Latavia	Food	87
<i>Lymnaea</i> sp.	Europe	Therapeutic uses	22
Thebapisana Otalalastasa	Portugal	Food	7
Otalalactaea	Portugal	Food	
	South America	- 1	00.00
Helix aspersa maxima	Brazil	Food	88,89

species include the snails Achatina sp., Archachatina marginata, Limicolaria sp., Lanistes varicus, Nucella lapillus, Lymnaea stagnalis, Helix sp., Pila sp., Semisulcospira sp., Pomacea canaliculata, Bellamya sp., Theba pisana and Otala lactea. Escargots (mainly Helix pomatia but some other species as well) are considered a delicacy in France and some other European countries. A systematic review of the literature reveals that the consumption of snails as a planned part of the diet is more common in Africa and Asia, although the acceptance varies to a great extent from region to region.

Although these species are used primarily as food, ethnomedicinal uses as reviewed by Meyer-Rochoware are also common and involve a wide variety of slugs and snails in connection with dermatoses, gastro-enteric troubles, coughs, colds, fractures²². ailments and even bone heart Consumption of snail soup or curry containing, for example, Bellamya sp., Pila sp., Lamellidens sp., or Parrevsia sp. is common among the people of the Kosi River basin of Bihar (India) for treating asthma. arthritis, rickets, cardiac disorders, giddiness, night blindness, etc.²³. An ethnozoological use of the garden snail (Helix aspersa) for the treatment of tuberculosis by the indigenous people of the Chindwara District of Madhya Pradesh (India)²⁴, the use of Pila sp. snails for treating weakness and paralysis by the tribal people and communities of the Gondwana Vindhyan region of Madhya Pradesh (India)²⁵ and the use of snails for bone strength, for treatment of body ache, joint pain, etc. by the Baiga people of Baihar tehsil of Balaghat district of Madhya Pradesh (India)¹⁴ are examples of therapeutic uses of snails in India.

Edible snails belong to the molluscan class Gastropoda and families like Achatinidae, Helicidae, Ampullariidae, Viviparidae, Thiaridae, Planorbidae, Pachychitidae, Turbinidae are represented. In general, a generic freshwater snail is dioecious, while those belonging to the subclass Pulmonata are hermaphrodites²⁶. Even in the case of the hermaphroditic species mating between two individuals occur followed by the deposition of eggs often covered with a protective gelatinous developmental coat. Although duration and life cycles are species-specific, under favourable conditions eggs usually hatch after about 2 weeks to one month. Thiarid (family Thiaridae) females parthenogenic²⁶ and Ampullarids are (family Ampullariidae), commonly known as 'apple snails',

are mostly amphibious with a mantle cavity containing organs that function as gills and lungs. *Pila* is a common and widely distributed genus in tropical Asia, but *Pomacea canaliculata* is an example that was introduced to many Asian countries from the USA and South America and became established^{11,27}. The Achatinidae is a family containing terrestrial pulmonate gastropods from Africa, such as the genera *Achatina*, *Limicolaria*, and *Archachatina*. Viviparidae are often known as river snails and contain livebearing species in which the females do not lay eggs, but retain them in their body.

Nutrient composition of edible snails

Moisture content

The moisture content of edible snail species varies within a range of 65.8 to 89.2% (Fig. 2 and Table 2). Fresh snails are generally used in preparations that include boiling, smoking, and using them for curries. In France, *Escargots-in-garlic-and-parsley-butter* is a famous snail dish; other worthwhile snail cuisines are the *Gaeng-Kui-Hoi-Khom* fresh water snail in red curry in Thailand, the *Tharoi Thongba* river snail curry in Manipur (India), the *Óchuong* stir-fried snail and *BúnÓc* in Vietnam, the *Lumachealla Romana* in Rome (Italy), *Cargols* in Spain, *Caracóis* stewed snail in Portugal, and *Kohlibourbouristi* pan-fried snail smakes their meat more susceptible to microbial contamination²⁸. Therefore in order to increase the



Fig. 2 — Proximate composition (% of fresh weight basis) of edible snail species. Number indicates the mean value of corresponding proximate composition i.e., moisture, protein, fat, ash [Graphical representation was plotted based on Table 2].

Scientific name		Proximate composition (% Fresh weight basis)							
	Moisture	Protein	Lipid	Fibre	Ash	NFE/Carbohydrate	-		
		Lar							
Achatina sp.	89.20	9.90	1.40		2.10	4.40	90		
Helix aspersa (small, 2.5 g)	83.30	12.00	0.70		2.70	0.50	91		
Helix aspersa (large, 3.5 g)	87.60	9.90	0.50		1.20	0.40			
Achatina fulica (Gray)	82.18	11.53	0.91		1.39		70		
Achatina fulica (Yellow)	82.36	13.42	1.28		1.29				
Achatina fulica (White)	81.20	13.69	0.97		1.25				
Limicolaria aurora [®]	71.24	14.78	2.79		3.38	7.79	51		
Archachatina marginata	76.63	20.76	1.20	ND	1.42		52		
Archatina sp.	79.89	14.52	4.24	ND	1.34				
<i>limicolaria</i> sp.	77.90	17.51	4.30	ND	1.12				
Helix aspersa aspersa	83.70	10.50	1.60		1.50		92		
Helix aspersa maxima	85.80	8.10	1.50		1.20	1.70			
Helix lucorum	84.00	10.80	1.50		1.40				
Helix pomatia	84.20	10.70	1.10		1.80				
Helix aspersa maxima***	79.85	9.50	2.66		0.92		89		
-	79.05	10.47	1.51		0.65				
	77.47	11.31	0.45		0.76				
	78.41	12.56	0.63		0.65				
Helix aspersa	80.80	16.35	0.41		1.89		74		
Archachatina marginata (ovum) Pfeiffer	76.56	20.56	1.38	ND	1.44	0.01	56		
Archachatina marginata (saturalis) Philippi	76.67	20.34	1.23	ND	1.40	0.37			
1 Achatina achatina	77.54	19.27	1.43	ND	1.34	0.42			
<i>imicolaria</i> sp.	78.68	18.66	1.17	ND	1.35	0.15			
Irchachatina marginata ovum [*]	80.78	16.23	0.86	0.10	0.77	1.26	57		
Irchachatina marginata saturalis [*]	80.30	15.95	0.79	0.20	0.89	1.89	0,		
<i>imicolaria</i> sp.*	78.64	15.33	0.80	0.21	1.50	3.31			
Irchachatina marginata	73.67	19.53	2.44		2.56	1.80	58		
Ichatina achatina	75.28	17.20	2.21		2.33	2.98	50		
Ichatina fulica	79.28	10.08	1.61		1.78	7.25			
<i>Limicolaria</i> sp.	84.91	5.86	1.01		1.78	6.91			
Helix pomatia	79.57	15.44	1.18	0.00	1.27		59		
Helix sp.	83.20	8.64	0.57	0.004	1.20	6.55	93		
Telix sp. Telix aspersa	83.20 82.50	8.04 12.87	0.57	0.004	1.04	4.99	93 79		
	82.30 73.95	20.22			1.73	1.09	65		
Archachatina marginata ^{**}			2.09	1.01			05		
	79.67	15.77	1.46	0.79	1.35	0.86			
	78.85	16.42	1.60	0.82	1.40	0.92			
	77.41	12.54	1.73	0.88	1.51	0.95			
	77.00	17.88	1.77	0.89	1.53	0.94	0.4		
Archachatina marginata (Adult)	79.25	17.22	1.46		1.34		94		
Archachatina marginata (Grower)	79.93	16.30	1.35		1.24				
Archachatina marginata (Snailet)	80.39	15.83	1.24		1.18		_		
Dtala lactea (boiled)	83.50	9.60	1.26		1.38	0.28	7		
<i>Otala lactea</i> (cooked)	78.60	12.90	1.54		3.45	0.32			
Theba pisana (boiled)	83.20	11.40	2.33		1.11	0.21			
Theba pisana (cooked)	77.50	13.40	3.23		3.05	0.35			
		Freshv	vater						
Pila globosa	85.50	8.27	0.73	0.03	2.60	2.90	93		
Sellamya bengalensis	82.10	8.97	0.98	0.04	3.64	4.31			
Ielaniatu berculata	74.60	12.36	1.79	0.05	3.68	7.57			
Inisus convexiusculus	75.70	12.93	0.97	0.04	4.61	5.79			
Brotia costula	69.86	12.91	0.82	ND	7.28	9.12	81		
Bellamya bengalensis	65.80	13.14	0.96	ND	8.11	11.97	Ŭ.		
Bellamya dissimilis	65.80	11.18	0.96	ND	8.15	11.46			
Pila globosa	73.80	15.59	1.15	ND	3.82	5.62			
	/2.00		1.10		2.02	5.02	(Cont		

	Table 2 —	- Proximate c	ompositior	of edible	e snail sp	ecies (Co	ntd.)	
Scientific name			Proxima	te compo	sition (%	Fresh we	ight basis)	Reference
		Moisture	Protein	Lipid	Fibre	Ash	NFE/Carbohydrate	
			Mari	ne				
Turbo militaris		73.08	16.19	5.57		2.14	3.02	85
Lunella undulata		70.83	18.49	5.20		1.97	3.51	
Lunella torquata		68.50	18.03	8.46		2.10	2.92	

*Calculated form obtained data on the dry weight basis to fresh weight basis

**Five values were obtained for the samples collected from five regions

**Four values were obtained for each component because of four different feed treatments

ND= Not determined



Fig. 3 — Comparative account of a) protein content (g/100 g fresh weight basis) of edible snails and conventional foods of animal origin; and b) fat content (g/100 g fresh weight basis) of edible snails and conventional foods of animal origin. [Data other than edible snails were obtained from USDA database https://fdc.nal.usda.gov accessed 10th February 2021].

shelf life of snails, drying prior to processing them further can be useful.

Protein and amino acids

The protein content of snail meat was found to be within the range of 5.9 to 20.8% with an average value of 14.0% on the basis of fresh matter (Fig. 2 and Table 2). The protein content of edible snails was therefore comparable to that of conventional animal meats (Fig. 3a). However, if converted on the dry weight basis using an average moisture content of 78.7%, the protein content reaches approximately 65.7%, which is higher than the protein contents of conventional protein sources. The nutritive quality of a protein depends on its amino acid composition. Fig. 4 represents the amino acid compositions of edible snails (Table 3). For several species, tryptophan was not determined presumably due to the acid hydrolysis process. Cysteine and methionine amounts are also generally not unambiguously identifiable due to the acid hydrolysis process in the analysis. Apart from the methodological concerns, the average proportion of essential amino acids was found to be about 36.5% while the nonessential amino acids accounted for 51.5%. Of course, the proportion will



Fig. 4 — Compositional distribution of amino acids on the basis of % of protein of edible snails. Number indicates the mean value of corresponding individual amino acid [Graphical representation was plotted based on Table 3].

be enhanced if cysteine and tyrosine are included as conditionally essential amino acids and arginine as essential for children. Among the essential amino acids, leucine (5.4-11.5%) and lysine (5.0-8.4%) are found predominant while glutamic acid (1-17.3%)

			Т	able .	3 — A	Amino	o acid	com	oositio	on of	edible	e snail	s						
Scientific name						Am	ino a	cid co	mpos	ition	(g/10	00 g protein)						Reference	
	Val	Leu	Lys	Ile	Thr	Met	Cys	Trp	His	Phe	Tyr	Ser	Arg	Pro	Asp	Glu	Gly	Ala	
Achatina fulica (Gray)	6.40	7.19	5.33	5.35	4.91	2.46	1.43	0.68	2.09	4.61	2.82	4.37	7.57	7.11	8.37	13.59	6.38	3.88	70
Achatina fulica (Yellow)	6.46	6.64	4.95	4.83	4.93	3.63	1.44	0.53	3.08	4.45	2.31	5.13	7.00	5.57	8.73	14.37	6.27	4.39	
Achatina fulica (White)	6.84	6.64	5.00	4.85	5.00	3.45	1.44	0.65	2.95	4.46	2.28	4.94	7.05	5.69	8.09	14.61	6.23	4.38	
Limicolaria aurora	7.60	11.50	7.20	7.30	3.90	1.40	0.90	ND	3.70	6.70	5.90	4.00	0.30	5.60	7.80	13.50	6.50	6.90	51
<i>Limicolaria</i> sp.	4.12	6.90	8.28	5.24	1.80	1.81	0.48	ND	3.56	4.37	2.58	4.05	6.74	3.99	7.61	1.01	5.06	3.98	54
Archatina archatina	3.28	5.58	6.06	4.46	2.24	1.37	0.87	ND	2.96	3.84	2.48	3.17	6.31	3.01	6.94	11.10	5.16	2.42	
Archachatina marginata	7.11	8.11	5.74	3.87	2.76	2.00	1.09	ND	4.70	4.73	2.93	3.33	5.96	3.61	7.27	14.4	4.57	5.19	
Hexaplex trunculus	4.72	6.96	8.35	4.81	1.93	1.99	0.36	ND	3.92	4.19	3.17	4.36	7.48	3.01	7.90	1.23	6.35	3.56	60
Helix aspersa [*]	6.34	5.42	6.39	4.12	4.00	3.78	3.47	ND	2.25	3.22	5.29	9.22	5.98	2.84	8.84	12.46	6.94	9.44	79
Cookia sulcata (small)	2.50	5.60	6.00	2.60	3.40	0.30	0.20	4.20	1.30	2.40	2.30	3.80	10.10	5.10	7.00	13.80	9.80	4.50	84
Cookia sulcata (big)	2.50	6.50	5.70	2.50	3.50	0.30	0.30	5.30	1.40	2.40	2.30	3.80	10.20	4.80	6.90	14.10	9.60	4.40	
Pomacea canaliculata	4.30	8.20	6.80	4.10	4.70	1.00	0.80	0.20	1.60	4.30	4.50	4.90	9.10	4.70	8.50	17.30	5.80	5.90	82
*Calculated based on obtain	ed value	s on th	ie basi	is of t	otal a	mino	acids	as pr	otein										
ND = Not determined																			

followed by aspartic acid (6.9-8.8%) was most abundant among the nonessential amino acids.

Currently, on average animal proteins contribute 25% of the protein intake worldwide, providing 18% of the calories²⁹. From the nutritional point of view, animal meat contains high-quality protein and highly bioavailable micronutrients like vitamin A, vitamin B12, riboflavin, iron, zinc, calcium, etc.³⁰. The consumption of meat in developed countries much higher than its consumption is in underdeveloped as well as developing regions of the world²⁹. However, the socio-economic changes like economic development, increased urbanization and globalization are leading to a transition in dietary patterns particularly in low and medium-income countries and increasing demand for animal protein is being observed³¹. At present, ruminants, chickens and pigs are contributing 96% to the global supply of animal protein, while aquaculture is growing fast³². However, not only the amount of protein available to the underprivileged societies is of concern, but to improve the production system and the distribution of food and at the same time guarantee sustainability is the challenge of the day. Forte et al., demonstrated that the snail carbon footprint amounted to 0.7 kg CO₂ eq. per kg of edible meat²⁰, which is far less than the amount generated by the conventional food sources of animal origin like chicken (1.8-5.2), pork (3.6-8.9), beef $(11-51)^{33}$, and even the newly advocated mealworm food source with its 2.7 kg CO₂ eq. per kg^{34} . Unlike cattle, the carbon footprint of snails is not affected by biogenic enteric methane emission but mainly derived from the feed crop cultivation and processing stages²⁰.



Fig. 5 — Distribution of different categories of fatty acids, i.e. Saturated fatty acids (SFA), Monounsaturated fatty acids (MUFA) and Polyunsaturated fatty acids (PUFA) on the basis of % of total fatty acids of edible snails. Number indicates the mean value of corresponding category of fatty acid [Graphical representation was plotted based on Table 4].

Fat and fatty acid composition

Fat content of snail meat was found to be within the range of 0.4 to 8.5% with an average value of 1.4% on the basis of fresh matter (Table 2). Fat content of edible snails was found lower than that of conventional foods of animal origin (Fig. 3b). Data on snail meat fatty acid profiles available from the literature have been compiled in Fig. 5 and Table 4. Based on their degree of saturation, fatty acids are categorized into three groups, i.e. saturated fatty acids (SFA), mono-unsaturated fatty acids (MUFA) and poly-unsaturated fatty acids (PUFA). With the exception of a few cases, the PUFA content of edible

-		PUFA	A (% of tota	al fatty aci	ds) of edi	ble snails				
Scientific name			Fatty	acid com	osition (%	6)				Reference
	Myristic	Palmitic	Stearic	Total	Oleic	Total	Linoleic	Linolenic	Total	-
	nijiistie	1 unintite	Stearre	SFA	01010	MUFA	(n-6)	(n-3)	PUFA	
Helix pomatia	0.42	10.29	16.38	37.87	14.70	19.65	13.56	1.87	25.83	74
Helix aspersa maxima	0.57	10.11	13.47	26.26	20.42	23.82	19.37	1.04	49.92	89
	0.42	7.68	14.51	24.86	18.61	22.68	19.74	1.10	52.46	
	0.52	8.11	12.32	23.06	18.32	21.63	24.53	1.34	55.31	
	0.65	6.72	12.59	22.20	16.73	20.74	22.70	1.32	57.06	
Helix aspersa	ND	7.28	17.37	28.76	13.57	20.66	17.53	5.63	34.38	79
Hexaplex trunculus	1.23	6.21	6.01	33.44	3.08	9.84	0.83	3.07	58.37	60
Melanopsis praemorsa [*]	5.66	20.04	8.37	39.00	10.36	21.30	16.33	8.87	39.98	95
	2.35	16.43	8.65	31.12	24.20	40.28	9.37	11.00	28.15	
	5.86	26.48	10.70	46.87	12.60	21.92	12.11	10.72	31.68	
	10.58	33.01	7.27	57.61	10.26	17.33	7.02	5.07	25.50	
Cookia sulcate (Small)	2.30	27.30	8.10	46.60	8.80	14.90	2.30	0.50	32.30	84
Cookia sulcate (Large)	2.30	2.40	9.60	42.20	8.00	10.80	2.10	1.00	36.30	
Assyriellaescheriana [†]	0.58	10.16	10.30	23.05	12.68	14.97	20.45	8.42	62.38	96
Assyriellaguttata [†]	1.85	9.61	12.33	26.95	15.06	16.14	18.54	6.24	56.80	20
Lunellatorquata	0.55	23.01	5.68	39.95	8.21	14.45	1.60	0.84	45.59	85
Lunellaundulata	0.96	21.62	6.59	38.61	8.41	14.83	2.82	2.33	46.58	00
Turbo militaris	0.13	22.12	5.45	39.83	7.89	14.05	2.92	1.87	45.13	
Pomacea canaliculata	3.10	20.50	9.00	39.50	9.10	17.70	20.60	0.90	42.70	82
Helix lucorum	0.53	9.66	6.75	21.70	22.02	26.53	21.76	10.08	51.77	97
Eobania vermiculata	0.30	10.49	7.76	22.68	19.07	21.77	37.67	7.31	55.56	
Helix pomatia (Farm A) ^{††}	0.47	10.16	14.31	34.84	13.83	19.55	17.95	2.47	39.68	98
(1 min 1)	0.52	11.24	15.51	37.98	15.57	19.91	16.95	1.57	34.14	20
	0.54	11.32	15.87	39.26	15.69	20.06	16.69	1.42	32.11	
<i>Helix pomatia</i> (Farm B) ^{††}	0.47	10.28	14.37	35.21	13.65	19.83	17.87	2.54	38.78	
	0.54	11.39	15.64	38.38	15.63	20.00	16.59	1.40	32.63	
	0.56	11.45	15.84	38.40	15.70	20.16	16.47	1.41	30.89	
Cornuaspersum maxima	0.34	8.36	12.18	28.72	20.36	24.22	32.42	3.50	44.58	
$(Farm A)^{\dagger\dagger}$	0.38	10.13	13.74	32.69	23.47	24.78	30.42	1.82	38.58	
	0.38	10.31	13.66	33.49	23.50	27.88	30.43	1.45	38.00	
Cornuaspersum maxima	0.35	8.27	12.23	28.32	26.64	30.91	27.42	3.70	39.81	
(Farm B) ^{††}	0.40	10.26	13.83	32.88	30.79	31.48	24.55	2.80	31.54	
	0.43	10.34	13.91	33.80	30.80	35.59	25.51	2.70	31.57	
Cornuaspersumaspersum	0.33	8.93	12.28	29.35	19.56	23.89	34.34	3.70	46.65	
$(Farm A)^{\dagger\dagger}$	0.37	10.17	13.87	32.87	23.18	24.25	31.84	2.22	40.11	
	0.38	10.5	13.96	33.92	23.40	28.13	31.71	2.21	39.34	
Cornuaspersumaspersum	0.35	8.66	12.89	29.54	25.81	29.85	27.40	3.85	41.30	
(Farm B) ^{††}	0.41	10.22	13.77	32.90	28.74	33.55	25.12	2.86	33.99	
	0.44	10.51	13.94	33.93	29.02	33.72	25.02	2.44	33.09	
Thebapisana ^{**}	0.73	12.63	5.41	20.72	28.83	31.27	18.78	7.64	48.10	99
· · · · r · · · · · · ·	1.06	15.75	6.31	25.32	28.83	30.49	21.35	8.87	44.19	
Cornuaspersum**	0.76	16.02	7.72	26.97	23.79	25.81	22.15	15.78	47.22	
4	0.59	13.24	7.24	23.01	29.95	33.53	19.07	15.14	43.56	
Eobania vermiculata**	0.81	14.63	7.66	25.58	26.03	27.86	21.94	12.40	46.56	
	0.63	13.31	7.59	23.35	29.71	32.24	19.20	15.53	44.41	
*										

 Table 4 — Fatty acid composition mentioning selective predominant fatty acids and three categories of fatty acids i.e. SFA, MUFA, PUFA (% of total fatty acids) of edible snails

*fatty acid content of the species was investigated in different seasons like autumn, winter, spring and summer (from the upper row downwards respectively)

[†]these species are not eaten

^{††}fatty acid profile was investigated in three forms i.e. raw, cooked and frozen (from upper row downwards respectively)

** the species was examined on the raw (upper row) and boiled (lower row) condition

snails is predominating and followed by SFA and MUFA contents. SFA content varied within the range of 20.7 to 57.6%, while MUFA and PUFA accounted for 9.8-40.3 and 25.5-62.4%, respectively. Statistical analysis (one-way ANOVA) with the available values for different categories of fatty acids reveals a significant difference among the three categories of fatty acids (df = 2, 132; F = 57.435, p = 0.000). Fatty acids play many different critical roles in the human body, being structural components of membrane lipids, precursors of molecules like eicosanoids, prostaglandins, and essential components of the metabolism. Saturated fatty acids, in general, are not desirable, because of their linkage with an elevation of cholesterols and thus cardiovascular disease. Lauric, myristic, and palmitic acids contribute comparatively more to a higher level of cholesterol³⁵, but stearic acid has been reported to lower the cholesterol level³⁶⁻³⁸. Among the SFAs, palmitic and stearic acid were the most abundant in the snail's fat, but oleic acid was the dominant component of the MUFAs. However, the distribution of the fatty acids belonging to PUFA category varied so widely that generalizations at this stage are not possible.

Mineral contents

Minerals contents of edible snails are shown in Fig. 6(a-b) and Table 5. Calcium, an essential macromineral, is found to be one of the most abundant minerals in edible snails. In humans, the majority of calcium by virtue of its phosphate salt is present in bones and teeth. It plays vital roles in several enzymemediated processes, excretion, blood clotting, and neuromuscular functions. Suboptimal intakes of calcium may hinder normal growth and can lead to manifestations of osteoporosis in older people, especially postmenopausal women³⁹. The high

potassium and low sodium content of edible snails should be expected to exhibit physiological benefits. Low potassium consumption is often associated with a variety of physiological disorders including respiratory, renal problems and hypertension. Further, edible snails are found to be rich in micro-minerals like iron and zinc. The most vulnerable sections of the population suffering from iron deficiency are infants of the weaning stage, young children, and women of childbearing age. More than 1.2 billion people are suffering from iron deficiency anaemia and iron deficiency in the absence of anaemia is even more frequent³⁸. Iron deficiency anaemia is often caused by the low dietary intake of iron and reduced intake of bio-available iron, particularly in developing countries⁴⁰. Zinc is an essential component of a large number of enzymes (more than 300). Moreover, it has anti-oxidative stress anti-inflammatory and functions⁴¹. Copper, together with zinc, are additional structural components of the antioxidant enzyme superoxide dismutase. Considerable amounts of magnesium, phosphorus, and manganese are also present in edible snails. However, to study the bioavailability of the individual minerals from the snail meat remains a task of the future.

Safety issues

Although edible snails are nutritionally comparable, if not superior to conventional foods of animal origin, proper care has to be taken in relation to safety issues of using snails as food. Accumulation of heavy metals in freshwater snails is one important issue. Lau *et al.*, reported that two aquatic snail *Brotia costula* and *Clithon* sp. tissues contained arsenic contents that were higher than the permissible limit for human consumption⁴². In another experiment on *Pomacea canaliculata* exposed to contaminate



Fig. 6 — Content of macro-minerals a) calcium, magnesium, sodium, potassium, and phosphorus (mg/100 g) and content of microminerals; and b) iron, zinc, copper and manganese content (mg/100 g) of edible snail species. [Graphical representation was plotted based on Table 5].

Scientific name	Mineral content (mg/100 g)											
	Ca	Mg	Na	K	<u>Р</u>	Fe	Zn	Cu	Mn	Reference		
Limicolaria aurora	401(DM)	771	178	533	636	ND	259			51		
Archachatina marginata	212.38 (FM)	46.33	196.24	44.18	0.64	9.31	1.44	0.64	0.41	52		
Archatina sp.	188.96	37.79	181.87	49.88	215.48	4.95	1.87	0.62	0.38	02		
Limicolaria sp.	22.24	18.91	53.83	72.82	183.35	4.66	1.23	0.16	0.59			
Helix aspersa (Foot)	200 (FM)	52.7			73	1	1	3.1		92		
liente aspersa (1 000)	1620 (DM)	425			590	8.4	8	25.2		2		
Helix aspersa (Viscera)	328	183			536	6.4	46.3	1.7				
(() isocial)	1120	625			1830	22	158	6				
Helix aspersa maxima	183	76			83	1.4	0.8	2.1				
Foot)	1390	575			630	11.2	6.4	16.4				
Helix aspersa maxima	356	182			473	10.2	43.1	1.2				
(Viscera)	1370	700			1820	39.2	166	4.8				
Helix lucorum (Foot)	279	60.8			66.8	0.9	1.3	3.4				
1011X 14001 4111 (1 001)	1840	400			440	6.0	8.8	22.8				
Helixlucorum (Viscera)	260	93.7			340	18	35.5	1.7				
1011111001 uni (v 15001a)	1040	375			1360	72	142	6.8				
Helix pomatia (Foot)	613	50.2			1300	1.9	1.1	4				
	4580	30.2 375			1049	1.9	8.8	30				
Helix pomatia (Viscera)	328	134			487	7.6	60	1				
Tenx pomunu (Viscera)	1340	550			1990	31.2	248	4.4				
Lymnaea stagnalis (Meat)		56.7	020	399		0.54	2 4 0	4.4 0.26		55		
	118.8 (DM) 201.08(FM)		82.8	399 192.78				0.20 ND	0.39	55 56		
Archachatina marginata ovum) Pfeiffer	. ,	45.59	50.80		123.43	8.69	1.54			50		
Archachatina marginata (saturalis) Philippi	207.53	45.34	52.93	209.95	123.23	9.41	1.69	ND	0.38			
Achatina achatina	204.63	46.15	60.94	193.74	131.38	9.43	1.76	ND	0.39			
<i>Limicolaria</i> sp.	208.75	45.99	65.10	197.57	153.89	9.46	1.51	ND	0.38			
Archachatina marginata	180.71(DM)	29.2		60.79	59.16	6.79	1.19			57		
ovum)												
Archachatina marginata	179.09	28.52		70	51.49	8.25	1.18					
saturalis)												
<i>Limicolaria</i> sp.	172.79	27.33		64.52	59.79	9	1.14					
Lanistes varicus	152.7	30.56		69.39	62.52	10.1	1.3					
Nucella lapillus	181.49	31.73		72.4	60.19	11.09	1.32					
Limicolaria sp.	36.2 (FM)	5.28			8.98	0.72		0.29		58		
Achatina fulica	66.3	15.13			14.79	1.3		0.58				
Achatina achatina	106.3	19.28			19.01	1.88		0.77				
Archachatina marginata	126.4	25.01			22.91	2.29		1.03				
Achatina fulica	186.35		28.97	64.15	62.73	1.51				59		
·	186.32		28.94	64.15	62.76	1.49						
<i>Limicolaria</i> sp.	179.65		24.78	56.72	58.34	1.36						
1	179.64		24.83	56.76	58.32	1.36						
Helix pomatia	13.51		13.63	31.59	31.17	1.17						
	132.48		13.65	31.57	31.14	1.14						
Hexaplex trunculus	674.4 (FM)	178.7	196.1	224.8	95.2	8.1	112.8	3.1	0.68	60		
Helix aspersa	135.70 (FM)	17.05	91.95	105.40	96.72	0.52				79		
Achatina achatina	585.5 (DM)			331.8	269.2	9.8	39	3.3		63		
Cookia sulcata (Small)	140 (DM)	330	400	300	110	9.8 5.81	3.22	0.64		84		
	70						3.22 3.82			04		
Cookia sulcata (Large)		310	410	280	90 550 5	7.41		0.78		02		
Pomacea canaliculata	5161.2 (DM)	56.9	93.4	364.4	550.5	45.5	10.1	7.1	2	82		
Lunella torquata	239 (FM)	69 (5	301	305	153	3.24	1.4	0.11	0.05	85		
Lunella undulata	44	65	270	333	164	4.11	1.52	0.06	0.05			
Turbo militaris	61	77	400	273	122	1.93	1.22	0.22	0.05			

	Та	ble 5 — Mir	neral content	t of edible s	nails (<i>Conte</i>	<i>d</i> .)								
Scientific name		Mineral content (mg/100 g)												
	Ca	Mg	Na	К	Р	Fe	Zn	Cu	Mn					
Limicolaria flammea	165.45	13.57	66.87	132.45	336.53	4.24	2.92	2.53	6.47	64				
Archachatina marginata (Adult)	42.19	59.23	44.75	92.34	295.64	9.53				94				
Archachatina marginata (Grower)	31.64	60.04	39.57	77.55	286.65	7.37								
Archachatina marginata (Snailet)	26.46	238.8	31.26	69.24	274.5	5.25								
ND = Not detected DM= Dry matter basis FM= Fresh weight basis														

sediment for two months it was revealed that higher than desired amounts of metals like iron, manganese, copper, and zinc had accumulated in the digestive tracts and glands⁴³. A similar trend was reported for Bellamva sp.⁴⁴. Onuoha et al., stated that snails of the species Archachatina marginata collected from the crude oil-producing regions in Rivers State, Nigeria were not safe for human consumption, because of the carcinogenic risk posed by the high amounts of nickel in them⁴⁵. Besides, edible snails can host a number of parasites^{28,46}. In a study by Tesana et al., from Northeast Thailand, the edible snail species Pila polita, P. pesmei, and Hemiplecta distincta all contained the nematode worm which Angiostrongylu scantonensis, can be transmitted to human hosts and cause Angiostrongyliasis⁴⁷. *Pomacea canaliculata* and Achatina fulica were found to be the most likely intermediate hosts of Angiostrongylus cantonensis in China⁴⁸. Among some other occasionally eaten snails, the species, Limicolaria aurora, L. flammea, and *Limicolariopsis* sp. were found to be contaminated with mesocercariae of the trematode Alaria, while L. flammea was furthermore found to contain cercariae of the liver fluke *Dicrocoelium dendriticum*⁴⁶. Therefore, in order to avoid these hazards, maintaining sanitation of the snail farm is necessary and proper processing of edible snails is also important⁴⁹.

Conclusion

Although snail farming is not a novel entrant in agriculture, it is far from receiving proper appreciation. Various species of snails are used for food and/or medicinal purposes, but their roles are primarily confined to within the limitation of a few users in a small number of countries. Often dietary transitions due to urbanization, availability of western foods, climate change, etc. are the major factors for the decline in the interest and the use of traditional foods including snails. To halt the erosion of the traditional food wisdom and to support the safe supply of snail meat, the establishment of helicicultural facilities is therefore, a necessity. An increase in snail meat consumption can mitigate nutrient deficiency and also enhance the economic condition of certain local areas in which suitable snails can be farmed and traded.

Acknowledgement

The study was partially supported by the BSRP grant through the National Research Foundation of Korea (NRF), Ministry of Education, grant number NRF-2018R1A6A1A03024862 received by Prof. Chuleui Jung.

Conflict of interest

The authors declare no conflict of interest.

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